© 2001 Wiley-Liss, Inc. genesis 30:157-159 (2001)

Morpholino-Induced Knockdown of *fgf8* Efficiently Phenocopies the *Acerebellar* (*Ace*) Phenotype

Isato Araki and Michael Brand*

Max-Planck-Institute for Molecular Cell Biology and Genetics, Dresden, Germany

Received 1 June 2001; Accepted 18 June 2001

Homozygous *acerebellar* (*ace*) embryos lack their cerebellum and the midbrain-hindbrain boundary (MHB) organizer, and in addition have defects in forebrain and heart development (Brand *et al.*, 1996; Picker *et al.*, 1999; Reifers *et al.*, 1998, Shanmugalingam et al. (2000); Raible and Brand, 2001; Araki and Brand, unpublished data), suggesting that this may be because of functional redundancy between Fgfs. Because morpholinos might help to resolve such issues, we sought to phenocopy the known defects caused by absence of Ace/Fgf8 through injection of a morpholino against *fgf8* (MO-*fgf8*). In spite of the complexity of the *ace* phenotype, we find that MO-*fgf8* efficiently and uniformly phenocopies the *ace* mutant in MHB, forebrain, and heart development.

We designed an antisense morpholino against fgf8, covering the translational start codon (Fig. 1J). After injections into one- to eight-cell-stage wild-type embryos, we find that MO-fgf8 effectively phenocopies the ace phenotype. We used between 0.5 and 4 µg/µl of MOfgf8 for the injection (Table 1), which delivers between 1.6 and 12.6 ng of MO-fgf8 per embryo. Morphologically, embryos at the 24-h stage injected with 1-4 μg/μl lacked the cerebellum and the MHB organizer, as do ace mutants at the same stage (Fig. 1A-I). A morpholino with four mismatched base pairs against fgf8 (control MO in Fig. 1J) had no effect, nor did it show any nonspecific effects at the same concentration (Table 1). To examine how closely the ace phenotype is mimicked, we stained the injected embryos with a probe for pax2.1 which reveals several of the tissues where Fgf8 functions. pax2.1 expression at the MHB is initially normal in ace mutants, but is not properly maintained (Reifers et al., 1998; Lun and Brand, 1998). At 24 h, MO-fgf8 injected embryos either lacked pax2.1 expression at the MHB completely, or it was reduced to a small dorsal patch, as seen in ace mutants at the same stage (Fig. 1D-I and not shown; Reifers et al., 1998). Because ace mutants show defects also in forebrain and heart development, we analyzed with molecular markers whether MO-fgf8 injection can phenocopy the *ace* mutant also in these tissues. Ten hours after fertilization, injected embryos have reduced and perturbed expression of emx1, an early telencephalic marker, as in ace mutants (Fig. 1K-M; Shanmugalingam et al., 2000). Similarly, at 24 h expression of pax2.1 in the optic stalk is reduced, and in the optic chiasm is reduced or missing (not shown), as found

previously for ace mutants (Shanmugalingam et al., 2000). Injected embryos stained with nkx2.5, an early marker for heart primordium, showed downregulation of nkx2.5, as is observed in ace mutants (Fig. 1N-P; Reifers et al., 2000a). We did not detect any nonspecific effect of MO-fgf8. We conclude that MO-fgf8 injection efficiently phenocopies the known loss-of-function phenotype of acerebellar in MHB, telencephalon, optic stalk, and heart development, thus validating the usefulness of this method. Because morpholino injection is thought to prevent translation (which we assume to be true, but have not tested, for fgf8), these findings also support (Reifers et al., 1998) that ace is a null allele. To further test this notion, we examined whether the phenotype of homozygous acerebellar mutants can be enhanced by Mo-fgf8 injection. Morphologically and after examining pax2.1 expression in injected embryos at the tailbud, two-somite, five-somite, and 24-h stage, we did not observe a difference between ace homozygotes and their wild-type siblings in the same clutch (Table 1 and not shown), showing that the ace phenotype cannot be further enhanced and therefore most likely represents the null phenotype. Morpholinos against other fgf mR-NAs alone or in combination with Mo-fgf8 will help to resolve the problem of redundancy that is often found with signaling by Fgfs.

ACKNOWLEDGMENTS

We thank Florian Raible and Steffen Scholpp for helpful comments, and Muriel Rhinn for technical advice. I.A. is supported by the Alexander von Humboldt Stiftung. This work was supported by grants from the Deutsche Forschungsgemeinschaft, the EU Biotech Program, and the Max-Planck Society to M.B.

LITERATURE CITED

Brand M, Heisenberg C-P, Jiang Y-J, Beuchle D, Lun K, van Eeden FJM, Furutani-Seiki M, Granato M, Haffter P, Hammerschmidt M, Kane

Contract grant sponsors: Alexander von Humboldt Stiftung, Deutsche Forschungsgemeinschaft, EU Biotech Program, and Max-Planck Society.

^{*} Correspondence to: Michael Brand, Max-Planck-Institute for Molecular Cell Biology and Genetics, Pfotenhauerstr 108, D-01307 Dresden, Germany. E-mail: brand@mpi-cbg.de

158 ARAKI AND BRAND

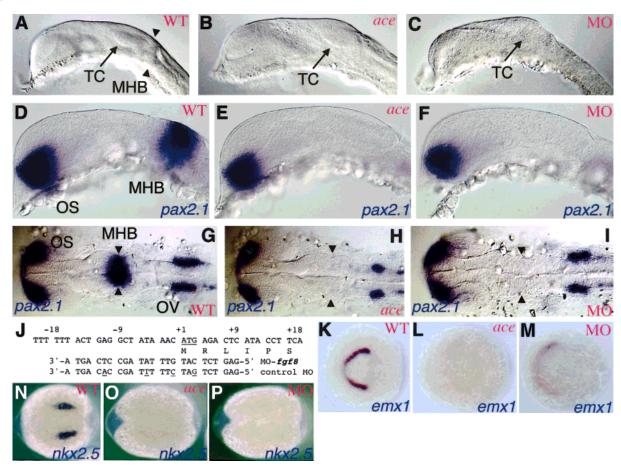


FIG. 1. Injection of a morpholino against fgf8 phenocopies the ace phenotype. (**A–K**) MO-fgf8 phenocopied ace phenotype in the midbrain-hindbrain boundary (MHB). Injected embryos or ace embryos lack the structure of MHB (arrowheads in A), the conspicuous constriction at MHB (compare arrowheads in G and H/l), and pax2.1 expression at MHB. The embryos were injected with MO-fgf8 at 2 μ g/ μ l in this figure. The embryos in A, D, and G are wild type; the embryos in B, E, and H are ace mutants. The embryos in D-I were stained with probe for pax2.1 mRNA. The pictures in A-F are lateral view, whereas the ones in G-I are dorsal view. In A-I, embryos were fixed at 24 h after fertilization. (**J**) The nucleotide/amino acid sequence around the initiation codon (underlined) of fgf8 cDNA and the sequences of the morpholino against fgf8 (MO-fgf8) and its negative control (control MO) are shown. The control-MO contains four single nucleotide exchanges (underlined) that are predicted to lead to a strongly reduced target binding (Gene Tools, LLC, Corvallis, OR). (**K-P**) MO-fgf8 phenocopied ace phenotype also in other tissues. The embryos in K and N are wild type; the embryos in L and O are ace mutant; the embryos in M and P are MO-fgf8-injected. The embryos were stained with an emx1 probe and a nkx2.5 probe in K-M and N-P, respectively. The pictures in K-P are dorsal views. Embryos in K-M were fixed at 10 h after fertilization; at 12 h in N-P. All embryos in this figure are oriented with rostral to the left. OS, optic stalk; OV, otic vesicle; TC, tectum. Methods: Crystallized morpholino oligos (Gene Tools, LLC) the injected/control embryos were raised at 28°C.

 Table 1

 Dose Response to a Morpholino Against Fgf8

Morpholino	Concentration (μg/μl)	Dose (ng)	n	pax2.1 staining at MHB		
				Absent	Reduced	Normal
MO-Fgf8	0.5	1.6	30		20 (67%)	9 (30%)
	1	3.1	103	50 (49%)	53 (51%)	O
	2	6.3	101	60 (59%)	41 (41%)	0
	4	12.6	24	10 (42%)	14 (58%)	0
	4	12.6	7	7 (100%)	`O	0
Injection into ace/ace homozygotes	2	6.3	63	58 (93%)	5 (8%)	0
Control MO	2	6.3	22	0	0	22 (100%)
	4	12.6	19	0	0	19 (100%)
Mock	0	0	23	0	0	23 (100%)

The readout was in situ hybridizations with pax2.1 probe at 24 h after fertilization.

- DA, Kelsh RN, Mullins MC, Odenthal J, Nüsslein-Volhard C. 1996. Mutations in zebrafish genes affecting the formation of the boundary between midbrain and hindbrain. Development 123:179–190.
- Lun K, Brand M. 1998. A series of *no isthmus (noi)* alleles of the zebrafish *pax2.1* gene reveals multiple signaling events in development of the midbrain-hindbrain boundary. Development 125: 3049–3062
- Picker A, Brennan C, Reifers F, Clarke JD, Holder N, Brand M. 1999. Requirement for the zebrafish mid-hindbrain boundary in midbrain polarisation, mapping and confinement of the retinotectal projection. Development 126:2967–2978.
- Raible F, Brand M. 2001. Tight transcriptional control of the ETS domain factors *Erm* and *Pea3* by FGF signaling during early zebrafish nervous system development. Mech Dev (in press).
- Reifers F, Böhli H, Walsh EC, Crossley PH, Stainier DYR, Brand M. 1998. *Fgf8* is mutated in zebrafish *acerebellar* (*ace*) mutants and is required for maintenance of midbrain-hindbrain boundary development and somitogenesis. Development 125:2381–2395.
- Reifers F, Walsh EC, Léger S, Stainier DYR, Brand M. 2000a. Induction and differentiation of the zebrafish heart requires fibroblast growth factor 8 (fgf8/acerebellar). Development 127:225-235.
- Reifers F, Adams J, Mason IJ, Schulte-Merker S, Brand M. 2000b. Overlapping and distinct functions provided by *Fgf17*, a new zebrafish member of the Fgf8/17/18 subgroup of Fgfs. Mech Dev 99:39–49.
- Shanmugalingam S, Houart C, Picker A, Reifers F, Macdonald R, Barth A, Griffin K, Brand M, Wilson SW. 2000. Ace/Fgf8 is required for forebrain commissure formation and patterning of the telencephalon. Development 127:2549-2561.