Natural product-inspired cascade synthesis yields modulators of centrosome integrity

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In biology-oriented synthesis, the scaffolds of biologically relevant compound classes inspire the synthesis of focused compound collections enriched in bioactivity. This criterion is, in particular, met by the scaffolds of natural products selected in evolution. The synthesis of natural product-inspired compound collections calls for efficient reaction sequences that preferably combine multiple individual transformations in one operation. Here we report the development of a one-pot, twelve-step cascade reaction sequence that includes nine different reactions and two opposing kinds of organocatalysis. The cascade sequence proceeds within 10-30 min and transforms readily available substrates into complex indoloquinolizines that resemble the core tetracyclic scaffold of numerous polycyclic indole alkaloids. Biological investigation of a corresponding focused compound collection revealed modulators of centrosome integrity, termed centrocountins, which caused fragmented and supernumerary centrosomes, chromosome congression defects, multipolar mitotic spindles, acentrosomal spindle poles and multipolar cell division by targeting the centrosome-associated proteins nucleophosmin and Crm1.

iology-oriented synthesis uses biological relevance as the key criterion for the synthesis of focused compound collections for chemical biology and medicinal chemistry research¹⁻⁴. The underlying scaffolds of natural product classes define the areas of chemical space explored by nature in evolution, and natural product-inspired compound collections¹ are expected to be biologically relevant and therefore enriched in biological activity⁵. Besides their role in drug discovery⁶, natural products have been instrumental⁷ in elucidating various biological processes related to cancer, particularly mitosis. The proteins involved in this highly regulated process seem to be particularly responsive to modulation by small molecules⁸⁻¹⁰; for instance, indole alkaloids have delivered many interesting candidates targeting mitosis (Fig. 1)^{11,12}. The structural complexity and diversity of natural products, and consequently also that of natural product-inspired compound collections, calls for the development of efficient synthesis methods that enable the flexible assembly of structurally diverse compound collections based on natural product scaffolds¹. Cascade reaction sequences in which several chemical reactions proceed consecutively in one pot can rapidly generate molecular complexity¹³⁻¹⁵. They require starting materials with multiple functional groups to facilitate different chemical transformations¹⁶. Here we report on the development of a highly efficient cascade reaction sequence of unprecedented length that rapidly proceeds via 12 consecutive chemical transformations in one pot, includes nine different chemical reactions (conjugate additions of P-, O-, N- and C-nucleophiles; acyclic and cyclic aminal formation; enamine condensation to form dihydropyridines; the aza-Claisen reaction; Pictet-Spengler cyclization; and chromone ring opening) and two opposing types of organocatalysis (Brønsted

acid and phosphine catalysis), and terminates in the formation of tetracyclic tetrahydroindolo[2,3-a]quinolizines, which resemble the core scaffold of numerous polycyclic indole alkaloids (**Fig. 1**)¹⁷. A corresponding focused compound collection yielded modulators of centrosome integrity that induce fragmented and supernumerary centrosomes, formation of multipolar mitotic spindles and acentrosomal spindle poles, defects in metaphase plate formation, multipolar cell division, and mitotic arrest and apoptosis, and that exert their biological activity by binding to the centrosome-associated proteins nucleophosmin (NPM1, which is also known as B23 or numatrin)¹⁸ and Crm1 (ref. 19).

RESULTS

Synthesis of a natural product-inspired compound collection Upon treatment with an organocatalyst-like triphenylphosphine, electron-poor 3-formyl chromones 1 and alkynes 2 undergo a

electron-poor 3-formyl chromones **1** and alkynes **2** undergo a [4 + 2] annulation reaction to yield natural product–inspired tricyclic benzopyrones **5**. The three-step, one-pot reaction proceeds via conjugate addition of the phosphine to the triple bond, conjugate addition of the resulting zwitterion enolate **3** to the chromone and subsequent annulation by conjugate addition of the oxygen anion to an α , β -unsaturated ester (**Scheme 1**)²⁰. The resulting tricyclic benzopyrones **5** incorporate two electrophilic α , β -unsaturated carbonyl groups and a pronucleophilic benzopyrone ring, which may open up to generate a nucleophilic phenol. Thus, if combined with reaction partners that also contain multiple functional groups, and if in the presence of suitable catalysts, they might serve as versatile starting points for new multistep reaction cascades. On the basis of this reasoning and with polycyclic indole alkaloid scaffolds in mind,

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we chose to combine the triphenylphosphine-catalyzed formation of tricyclic benzopyrones with tryptamine derivatives in the presence of an acid.

We envisioned that under suitable reaction conditions both natural product-based fragments equipped with more than one site of complementary reactivity could establish a cascade reaction that might yield unique natural product–inspired compound classes in a concise and efficient manner (**Fig. 1b**). In particular, it was considered possible that under suitable conditions the cascade sequence might lead to the formation of polycyclic indole derivatives such as indoloquinolizines and related heterocycles. The indoloquinolizine scaffold is characteristic of numerous polycyclic monoterpene



Scheme 1 | Cascade synthesis of indoloquinolizines 16a-16z using easily accessible substrates. Further details are in the text.

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Figure 1 | Inspiration and synthesis design for indoloquinolizine compound collections. (a) Indole alkaloid natural products that have diverse biological activities. (b) A cascade reaction-based retrosynthetic analysis suggests using tryptamine and benzopyrones as cascade substrates to yield indologuinolizines.

indole alkaloids with diverse biological activities (for example, yohimbine, hirsutine or augustin; Fig. 1a).

For reaction development, benzopyrone 5a was treated with tryptamine 6 for a few minutes at 80 °C followed by addition of camphorsulfonic acid, and the reaction mixture was stirred for an additional 30 min. Regioselective S_N2'-type addition to the less substituted α,β -unsaturated vinylogous ester would result in chromone ring opening to yield aminal 7. Conjugate addition of the intermediary phenol with concomitant retro-Michael pyran ring opening would lead to enamine 8. The subsequent nucleophilic attack of the enamine on the neighboring α -ketoester was expected to result in formation of dihydropyridine 9. Ring opening of chromones²¹ was expected to lead to the formation of a pyridinium intermediate 10, which then might have been trapped by the electron-rich indole to terminate the cascade, yielding indoloquinolizine 11. However, compound 16a was isolated, and its structure was unambiguously proven by crystal structure analysis (Supplementary Methods). Insight into the cascade reaction sequence emerged from the successful isolation of intermediates 12a and 12b (an X-ray crystal structure of 12b and spectroscopic characteristics of 12a are shown in Supplementary Methods).

We presume that the intermediates 12 are formed from 10 by addition of the phenol to the pyridinium ion and undergo subsequent [3,3]-sigmatropic aza-Claisen ring opening to yield α -imino esters 13, which are subject to acid-catalyzed Pictet-Spengler cyclization. The resulting secondary amines 14 then may form hexacyclic compounds 15 by another conjugate addition. Chromone ring opening, with the phenol serving as a good leaving group, finally results in the formation of the tetrahydroindolo[2,3-a]quinolizines 16. Under acidic conditions, indole rings do not add to the pyridinium cations^{22,23}, and therefore formation of 16 or 11 directly from 10 is very unlikely. We assume that the ester group ($R^4 = CO_2R$) attached to the enamine moiety in 12a lowers its lowest unoccupied molecular orbital and therefore facilitates the aza-Claisen rearrangement to yield imino-esters 13, which then enter the further cascade reaction sequence (steps 10 to 12) that terminates in indoloquinolizines 16. Further support for the proposed cascade mechanism was obtained from a separate experiment in which 12a was converted almost quantitatively to 16a upon exposure to acid, whereas 12b remained unchanged under these conditions (Scheme 1 and Supplementary Methods). Although these results lend very strong support to our mechanistic proposal, we note that an alternative plausible formation of 16a via a different, more direct mechanism cannot be completely ruled out.

This acid-catalyzed conversion of **5** into **16** in one pot could successfully be combined with the phosphine-catalyzed formation of **5** from **1**. The desired indoloquinolizines **16a–16z** are formed within 10–30 min (**Table 1**) in an efficient one-pot procedure by treating a mixture of **1**, **2** and triphenylphosphine at 80 °C in toluene with tryptamine (slow addition, 2–5 min) followed by addition of camphorsulfonic acid. The results given in **Table 1** show that the reaction is widely applicable and is notably high yielding. Using exclusively commercially available starting materials, the entire one-pot cascade reaction sequence proceeds via twelve different reaction steps, including nine different chemical transformations and two opposing types of catalysis. To the best of our knowledge, this is the longest cascade reaction sequence known^{13–15} The resulting heterocycles resemble the core scaffold structure of numerous alkaloids such as the yohimbines¹².

Investigation of mitosis modulation

Given the responsiveness of mitosis to small molecules, in particular indole alkaloids^{8,24}, we then tested our focused collection of indoloquinolizines for possible modulation of cell division using a phenotypic screen in BSC-1 cells³. Out of all of the tested compounds, **16a** led to the accumulation of round-shaped cells with condensed DNA, characteristics that are indicative of mitotic cells (**Supplementary Results** and **Supplementary Fig. 1**). Moreover, the mitotic cells showed impaired mitotic spindles.

Treatment of HeLa cells overexpressing GFP-tubulin and U2OS cells overexpressing mCherry- α -tubulin with **16a** and analogs there of induced multipolar cell division and abundant formation of three daughter cells during mitosis (Supplementary Movies 1 and 2). Impairment of mitosis in HeLa cells (Fig. 2 and Supplementary Fig. 2) and other cell lines (Supplementary Fig. 3) is dependent on both the concentration and structure of the compound, with 16a and 16h being the most potent compounds (mitotic defects can already be detected at a concentration of 1.5 µM; detailed in Supplementary Fig. 4 and Supplementary Results) and (R)-16a and (R)-16h being more potent than (S)-16a and (S)-16h (Supplementary Results and Supplementary Fig. 4; determination of the absolute configuration of 16a is detailed in Supplementary Methods). The substitution pattern in the aromatic ketone substructure and in the quinolizine cannot be changed substantially without loss of activity (Table 1). However, variation of R⁵ in the phenyl ring of the indole was tolerated (Table 1, entries 8, 22, 23 and 25).

(*R*)-16a and (*R*)-16h induced chromosome congression defects and tri- or even multipolarization of the mitotic spindle (Supplementary Fig. 4 and Supplementary Movies 3 and 4). In the parental HeLa cell line, centrosomes were visualized using γ -tubulin-specific antibody, and centrioles were stained with cep135-specific antibody. Inspection of the images shown in Figure 2a and Supplementary Figure 2 revealed the formation of different spindle poles that were either stained for both γ -tubulin and cep135, contained only one of both markers or represented acentrosomal poles. In addition, the number of stained centroles.





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Entry	Product	R	R ¹	R ²	R ³	R⁵	R ⁶	Percentage yield	Activity ^a
1	16a	Me	Н	Н	Н	Н	Н	58	++
2	16b	Et	Н	Н	Н	Н	Н	88	-
3	16c	Et	Me	Н	Н	Н	Н	65	-
4	16d	Me	Me	Н	Н	Н	Н	56	-
5	16e	Me	Br	Н	Н	OMe	Н	62	-
6	16f	Me	CI	Н	Н	Н	Н	39	-
7	16g	Me	Br	Н	Н	Н	Н	39	-
8	16h	Me	Н	Н	Н	OMe	Н	76	++
9	16 i	Me	ⁱ Pr	Н	Н	Н	Н	66	-
10	16j	Me	CI	Н	CI	Н	Н	20	-
11	16k	Me	Br	Н	Br	Н	Н	20	-
12	161	Me	CI	Me	Н	Н	Н	60	-
13	16m	Et	Н	Н	Н	OMe	Н	76	-
14	16n	Et	Br	Н	Н	Н	Н	52	-
15	160	Et	Me	Н	Н	OMe	Н	45	-
16	16p	Et	ⁱ Pr	Н	Н	OMe	Н	65	-
17	16q	Et	CI	Me	Н	Н	Н	40	-
18	16r	Et	CI	Н	Н	Н	Н	65	-
19	16s	Et	CI	Н	Н	OMe	Н	63	-
20	16t	Et	Br	Н	Н	OMe	Н	54	-
21	16u	Me	Phenyl	Phenyl	Н	Н	Н	52	-
22	16v	Me	Н	Н	Н	Br	Н	73⁵	+
23	16w	Me	Н	Н	Н	Me	Н	74 ^b	+
24	16x	Me	Н	OBn	Me	Н	Н	59	-
25	16y	Me	Н	Н	Н	ОН	Н	67°	+
26	16z	Me	Н	Н	Н	Н	CO ₂ Me	91 ^{b,c}	-

^{*}A qualitative phenotypic analysis using HeLa cells treated with 25 μ M of 16a-z was done, and molecules causing chromosomal misalignments were considered active ones (++, >90 % of all mitotic cells; +, > 30% of all mitotic cells; -, <10% of all mitotic cells). ^bThe reaction was carried out from the tricyclic benzopyrone **5** (**Supplementary Methods**, general procedure 2). ^cThe product was obtained as mixture of isomers. ⁱPr, isopropyl; OBn, O-benzyl.

These findings indicate that centrosomes may have been fragmented or that supernumerary centrosomes were formed.

Indoloquinolizines (*R*)-16a and (*R*)-16h inhibit cell proliferation, induce apoptosis as monitored by caspase 3 and caspase 7 activation (Supplementary Figs. 3b and 5 and Supplementary Table 1) and induce a pronounced mitotic delay in HeLa GFP-H2B cells (Supplementary Fig. 6) with cell cycle arrest at the M stage (Fig. 2b,c and Supplementary Fig. 7).

Identification of cellular targets

On the basis of this structure-activity relationship, we synthesized indoloquinolizine probes 17 and 18 (negative control) (Fig. 3a) for target identification by means of a chemical proteomics approach. Compound 17 was immobilized on Sepharose beads and exposed to HeLa cell lysates, and bound proteins were released by elution with a ten-fold excess of 16a followed by MS determination of the bound proteins, revealing that probe 17, but not control probe 18, binds the centrosomal protein NPM and U2 small ribonuclear protein (Supplementary Fig. 8).

Knockdown of NPM by small interfering RNA induces fragmentation of centrosomes and impairs chromosome congression and mitotic spindle formation in HeLa cells²⁵. NPM is involved in the regulation of centrosome duplication during mitosis and also promotes ribosome biogenesis, such that U2 small ribonuclear protein could be pulled down in complex with NPM. On the basis of these data, NPM was validated as a target protein. Reversible binding of NPM to 17 was confirmed by western blotting with an NPM-specific antibody and by concentration-dependent competition between immobilized and nonimmobilized ligand (Fig. 3b). Regulation of centrosome duplication by NPM includes complex formation with the nuclear export receptor Crm1 (ref. 26). Crm1 knockdown as well as Crm1 inhibition by leptomycin B leads to defects in chromosome alignment and spindle assembly^{26,27}. Reinvestigation of the affinity pull-down experiment by immunoblotting with a Crm1-specific antibody indeed revealed reversible, competitive binding of Crm1 to immobilized ligand 17 (Fig. 3b). Binding of both NPM and Crm1 was confirmed by means of fluorescence polarization measurements using fluorescent probe **19** (Fig. 3a), which revealed an apparent K_d of 25.4 ± 1.9 μ M for histidine-tagged NPM (His-NPM) and 8.8 \pm 1.6 μ M for Crm1 (Fig. 3c and Supplementary Fig. 10). Direct interaction between indoloquinolizine probe 19 and NPM as well as Crm1 in HeLa cells

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including chromosome congression defects (middle row) and aberrant spindle structures (bottom row) induced by (R)-16a. HeLa cells were treated with 25 μ M (R)- **16a** for 18 h before staining with antibodies specific for α -tubulin, γ -tubulin, cep135 and DNA. Images are Z projections of image stacks done with ImageJ software. Representative images of mitotic cells are shown for cells treated with DMSO (top) and cells treated with 25 µM (R)- 16a (middle and bottom). y-Tubulin staining is indicative of centrosomes, and cep135-staining is indicative of centrioles. Scale bar, 10 um. (b.c) FACS analysis of HeLa cells treated with (R)- 16a. Cells were incubated for 18 h with different concentrations of (R)- 16a and DMSO (as a control) before staining with phosphohistone 3-specific antibody. Alexa Fluor 488-labeled secondary antibody and propidium iodide. Representative histograms for DMSO and 7.5 uM (R)-16a (b) and dose-dependent accumulation of mitotic cells are shown (c).

was proven by means of a significant (P < 0.05) decrease in the fluorescence lifetime of the donors NPM-citrine and enhanced yellow fluorescent protein (EYFP)-Crm1 after addition of the Cy3labeled acceptor indoloquinolizine 19 (Fig. 3d,e). These findings reveal that both NPM and Crm1 independently bind the derivative 19. Nuclear export mediated by Crm1 can be monitored by following the cellular localization of RanBP1, a cargo molecule for Crm1 (ref. 28). Whereas HeLa cells treated with the known nuclear export inhibitor leptomycin B showed accumulation of RanBP1 in the nucleus, (R)-16a did not influence the cellular distribution of RanBP1 up to a concentration of 25 µM (Supplementary Fig. 11). At 50 μ M (*R*)-**16a**, the observed partial inhibition of nuclear export

b

demonstrated that the indologuinolizine targets Crm1. Notably, the indologuinolizines are much less cytotoxic than leptomycin B (in cell viability assays, half-maximal inhibitory concentration (IC₅₀) values for centrocountins are in the micromolar range, whereas leptomycin B has IC₅₀ values in the low nanomolar range²⁹; Supplementary Fig. 12). These results indicate that the indologuinolizines have a different mode of action than leptomycin B.

Our findings demonstrate that compound (R)-16a independently targets the centrosome-associated proteins NPM and Crm1 in cells. It thereby leads to impairment of centrosome and spindle integrity, chromosome congression defects, cell cycle arrest at the M stage and apoptosis.

e

d



Figure 3 | Identification and validation of NPM and Crm1 as target proteins of 17. (a) Structure of probes used in affinity pull-down (PD) experiments, fluorescence lifetime experiments and determination of fluorescence polarization. (b) Immunodetection of NPM and Crm1 binding to immobilized 17. After pull-down using immobilized 17, proteins released by heating were resolved by SDS-PAGE, transferred to a polyvinylidene fluoride membrane and subsequently detected with NPM- and Crm1-specific antibodies. For competition experiments, HeLa cell lysates were preincubated with different concentrations of (R)- 16a. Full-length blots are presented in Supplementary Figure 9. (c) Binding of 19 to NPM and Crm1 as determined by means of fluorescence polarization. The Cy3-labeled probe 19 was titrated with increasing concentrations of His-tagged NPM or Crm1 until saturation was reached. K_d values were determined from the fit of fluorescence polarization versus His-NPM or Crm1 concentration. (d) Fluorescence lifetime imaging microscopy showing specific binding of NPM-citrine and EYFP-Crm1 (donor) to Cy3-labeled 19 (acceptor). HeLa cells were transfected with NPM-citrine or EYFP-Crm1. Twenty-four hours later, 2.5 μ M 19 was added. The decrease in the lifetime of the donor after addition of 19 is shown in the lifetime maps of cells. Scale bars, 30 µm. (e) Graph showing the decrease in the donor lifetime as compared to the control. Data are expressed as mean $(n > 15) \pm$ s.d. *, **P < 0.05. P values were determined following two-tailed Student's t-test. A P value of < 0.05 was considered significant.

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DISCUSSION

We demonstrate the development of an exceptionally versatile reaction cascade of unprecedented length and efficiency, giving access to a structurally complex natural product-inspired compound collection that yields new modulators of centrosome integrity and mitotic spindle pole formation by simultaneously targeting NPM and Crm1. On the basis of the induced phenotype, we term these molecules centrocountins, with compound 16a being centrocountin 1. Although for chemical-biological analysis of Crm1 function leptomycin B is a widely used molecular probe³⁰, small molecules influencing NPM function have rarely been described, with the electrophilic and thereby nonselective natural product avrainvillamide being the most prominent case³¹. A dual NPM-Crm1 ligand is unique, and a molecular probe for the elucidation of NPM or NPM-Crm1 function is not available yet. NPM is involved in the establishment of many cancers, although its role is controversial³². In light of the efficient accessibility of the centrocountins and the fact that treatment of cancer cell lines with these compounds is antiproliferative and leads to mitotic arrest and apoptosis, the centrocountins and modulation of the activity of NPM or the NPM-Crm1 complex might inspire new drug discovery programs.

METHODS

General procedure for the cascade synthesis of indoloquinolizines 16. The 3-formylchromone (1 eq.) was dissolved in toluene (10 ml mmol⁻¹) by heating to 80 °C, and the acetylenedicarboxylate (1.3 eq.) and triphenylphosphine (0.6 eq.) were added. After 2–5 min, the tryptamine (1.1 eq.) was added, and, after the tryptamine dissolved, camphorsulfonic acid (1.5 eq.) was added. After 5–30 min, the solvent was evaporated, and the remaining residue was subjected to column chromatography on silica gel to yield the indoloquinolizine 16 as a yellow solid, which can be further purified by precipitation from methanol. Further details are available in **Supplementary Methods**.

Other methods. Cell lines and plasmids, imaging protocols, isolation of binding proteins and other cell-based procedures are detailed in **Supplementary Methods**.

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Author contributions

H.D., V.K. and H.B. designed and performed the synthesis experiments. V.P., S.M., A.W.B., Z.M. and S.Z. carried out the biological studies. P.J. and A.B. performed MS analysis. H.W., K.K., K.H., S.Z. and A.H. designed experiments. M.S. and H.P. carried out the X-ray crystallographic analysis. S.G. determined the absolute configuration of **16a**. H.W., K.K. and S.Z. supervised the project and wrote the manuscript. All authors discussed the results and commented on the manuscript.

Competing financial interests

The authors declare no competing financial interests.

Additional information

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