Synthesis and Hormonal Activity of the (25S)-Cholesten-26-oic Acids – Potent Ligands for the DAF-12 Receptor in *Caenorhabditis elegans*

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Using a highly stereoselective Evans aldol reaction for the introduction of the stereogenic center at C-25, we describe an efficient synthesis of the orthogonally diprotected (25*S*)-26-hydroxycholesterol **11**. In a few synthetic steps, this crucial intermediate **11** has been converted into the four (25*S*)-cholesten-26-oic acids **1–4**, which have been obtained in 12–15 steps and 19–53 % overall yield based on commercially available 3 β -hydroxychol-5-en-24-oic acid (**5**). Our biological studies of the compounds **1–4** reveal that (25*S*)- Δ^7 -dafach-

ronic acid (1) represents the most active steroidal ligand for the hormonal receptor DAF-12 in *Caenorhabditis elegans*. Moreover, the saturated (25*S*)-dafachronic acid (3) represents a new ligand for this receptor and the (25*S*)-steroidal acids are more active as compared to their corresponding (25*R*)counterparts.

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Introduction

Reproductive development of nematodes such as *Caenorhabditis elegans* and *Pristionchus pacificus* is controlled by steroidal ligands, called dafachronic acids (Figure 1).^[1,2] In *C. elegans*, the biosynthesis of these steroids requires activity of the cytochrome P450 DAF-9.^[3] Dafachronic acids are ligands which inactivate the nuclear hormone receptor DAF-12 and thus, lead to reproductive development of worms. In *daf-9* mutant worms, incapable of dafachronic acid biosynthesis, DAF-12 is active and worms enter the diapause state generating dauer larvae. Another ligand known to bind at DAF-12 is (25*S*)-cholestenoic acid (4).^[4]

Mangelsdorf and colleagues prepared (25S)- Δ^4 -dafachronic acid (2) and its C-25 epimer from the corresponding 26-hydroxycholesterols.^[1a] In 2007, the structure of the other ligand, (25S)- Δ^7 -dafachronic acid (1), has been confirmed by a synthesis from Corey and Giroux.^[5] Moreover, it has been shown that (25S)- Δ^7 -dafachronic acid (1) represents the most active ligand known so far.^[1a,5] Interestingly, the synthesis of both C-25 epimers of 2 and 4 was described previously by Khripach et al.^[6] Our investigations on the synthesis and biological activity of cholesterol derivatives,^[7] led us to an elegant and concise synthesis of the 25*R*-diastereoisomers of 1, 2 and 4 starting from commercially

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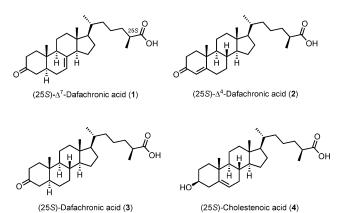


Figure 1. Hormonally active steroidal acids 1–4.

available diosgenin.^[8,9] Also in the 25*R*-series, the Δ^7 dafachronic acid exhibited the highest hormonal activity.^[9] Since yamogenin, the C-25 epimer of diosgenin, was not available from commercial sources, we devised a novel stereoselective construction of the side chain for the synthesis of all three (25*S*)-cholesten-26-oic acids **1**, **2** and **4**, as well as the saturated (25*S*)-dafachronic acid (**3**).^[10] The synthesis of (25*S*)-dafachronic acid (**3**) was also reported by Corey and co-workers.^[11]

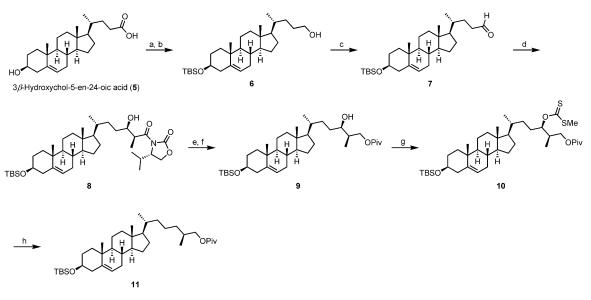
Results and Discussion

The Evans aldol reaction represents a powerful synthetic method for the enantioselective construction of stereogenic carbon centers.^[12,13] However, applications to stereoselective synthesis of steroid side chains are rare.^[14] For our pur-



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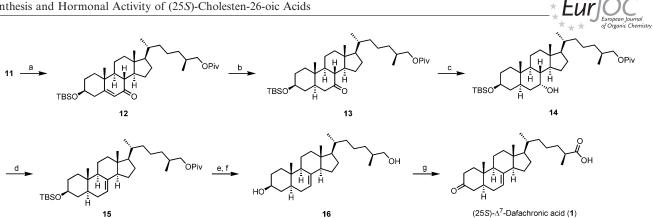
Scheme 1. Stereoselective synthesis of the crucial intermediate **11**. *Reagents and conditions:* (a) 3.2 equiv. TBSCl, 8.1 equiv. imidazole, 2.2 equiv. DMAP, DMF, 25 °C, 18 h; (b) 4.0 equiv. LiAlH₄, THF, 25 °C, 17 h, 96% for 2 steps; (c) 4.0 equiv. DMSO, 2.0 equiv. (COCl)₂, CH₂Cl₂, -78 °C, then **6**, 20 min, 5.0 equiv. Et₃N, to 25 °C, 98%; (d) 1.3 equiv. (*S*)-(+)-4-isopropyl-3-propionyl-2-oxazolidinone, 1.43 equiv. Bu₂BOTf, 1.69 equiv. Et₃N, CH₂Cl₂, 0 °C, 1 h, then -78 °C, 7, 30 min, then 0 °C, 80 min, 95%; (e) 1.1 equiv. LiBH₄, 1.0 equiv. H₂O, Et₂O, 0 °C, 2 h; (f) 1.3 equiv. PivCl, pyridine/CH₂Cl₂ (1:1), 0 °C, 2 h, 81% for 2 steps; (g) 1.0 equiv. NaHMDS, 20.0 equiv. CS₂, THF, -78 °C to 0 °C, then 2.0 equiv. MeI, 30 min, 98%; (h) 10 mol-% AIBN, 15.0 equiv. Bu₃SnH, reflux, 5 min, 93%.

poses, commercially available 3 β -hydroxychol-5-en-24-oic acid (**5**) appeared to be the ideal starting material (Scheme 1).^[15] Treatment of **5** with *tert*-butylchlorodime-thylsilane (TBSCI) in the presence of imidazole and DMAP led to the intermediate silyl ester which on subsequent reduction using lithium aluminium hydride provided almost quantitatively the 24-hydroxy derivative **6**. Oxidation of the alcohol **6** with PDC afforded the aldehyde **7** in 96% yield.

Moreover, Swern oxidation of 6 provided the aldehyde 7 in 98% yield even on large scale.^[16] By using the standard conditions reported by Evans.^[12] (1.3 equiv. of commercial (S)-(+)-4-isopropyl-3-propionyl-2-oxazolidinone, triethylamine and dibutylboron triflate), the aldol product 8 was available on large scale as a single stereoisomer in 95% yield. Reduction of 8 with lithium borohydride followed by selective pivaloylation of the primary hydroxy group afforded compound 9 in 81% yield over both steps. In the next step, the hydroxy group at C-24 had to be removed. Mesylation of the hydroxy group followed by treatment with lithium aluminium hydride afforded the corresponding C-26 hydroxy compound along with the C-24/C-25 olefin. Both compounds were not separable by flash chromatography on silica gel. The reaction of xanthates with tributylstannane in the presence of AIBN provides deoxygenated products in high yields and thus, represents a promising alternative.^[17] For the synthesis of the corresponding xanthate, alcohol 9 was treated with NaHMDS and carbon disulfide at low temperature. Addition of iodomethane provided the xanthate 10 almost quantitatively. After a reaction time of only 5 min, Barton deoxygenation of 10 afforded the deoxygenated compound 11 in 93% yield. In order to achieve good results in this transformation, recrystallisation of commercial AIBN from methanol is recommended. Using the strategy described above, the orthogonally diprotected (25S)-26-hydroxycholesterol 11 is readily available in 8 steps and 66% overall yield. Compound 11 represents the central intermediate of our synthesis and has been converted into the four (25S)-steroidal acids 1–4.

For an access to (25S)- Δ^7 -dafachronic acid (1), a double bond shift was required. Allylic oxidation following Chandrasekaran's procedure afforded the enone 12 in 75% yield (Scheme 2).^[18] Palladium-catalyzed hydrogenation using ethyl acetate as solvent led to the ketone 13.

The reduction of cyclohexanones using sterically hindered reducing agents under kinetic conditions provides the axial alcohols.^[19] Thus, treatment of ketone 13 with L-Selectride[®] afforded stereoselectively the 7 α -alcohol 14 in 90% yield. Using the commercially available Burgess reagent in benzene under reflux, the cholest-7-ene 15 could be isolated in 78% yield (Table 1).^[20] In the 25*R*-series, elimination of a structurally related alcohol with thionyl chloride in pyridine provided quantitatively the corresponding Δ^7 -olefin.^[8,9] Treatment of 14 with 3.0 equiv. of thionyl chloride in pyridine provided the olefin 15 in only 72% yield. However, increasing the amount of thionyl chloride to 5.0 equiv. afforded 15 in 87% yield. Removal of the silvl and pivaloyl protecting groups was achieved by treatment with first TBAF and then lithium aluminium hydride to afford the diol 16 in 82% yield. Changing the sequence of the removal of the protecting groups resulted in only 69% yield of 16. At this stage of our synthesis, the 25S-configuration of the 26-hydroxycholest-7-en-3 β -ol (16) has been unambiguously confirmed by an X-ray crystal structure determination (Figure 2).^[10] Finally, Jones oxidation of the diol 16 provided (25S)- Δ^7 -dafachronic acid (1) in 89% yield.



Scheme 2. Synthesis of (25S)- Δ^7 -dafachronic acid (1) from the intermediate 11. Reagents and conditions: (a) 4.0 equiv. PDC, 8.0 equiv. tBuOOH, Celite[®], benzene, 0 °C to 25 °C, 41 h, 75%; (b) 10% Pd/C, H₂, EtOAc, 25 °C, 16 h, 95%; (c) 1.3 equiv. L-Selectride[®], THF, -78 °C, 1.5 h, 90%; (d) 5.0 equiv. SOCl₂, pyridine, 0 °C, 40 min, 87%; (e) 1.5 equiv. TBAF, THF, reflux, 16 h; (f) 4.0 equiv. LiAlH₄, THF, 0 °C to 25 °C, 17 h, 82% for 2 steps; (g) 5.0 equiv. Jones reagent, acetone, 0 °C, 90 min, 89%.

Table 1. Elimination of the 7α -hydroxy group from 14.

Reaction conditions	% Yield of 15
2.0 equiv. Burgess reagent, benzene, reflux, 2 h	78
3.0 equiv. SOCl ₂ , pyridine, 0 °C, 30 min	72
5.0 equiv. SOCl ₂ , pyridine, 0 °C, 40 min	87

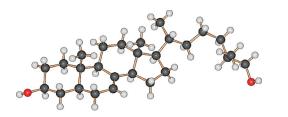


Figure 2. X-ray crystal structure of the diol 16 (orthorhombic, $P2_12_12_1).$

Transformation of intermediate 11 to (25S)- Δ^4 -dafachronic acid (2) required only a few steps (Scheme 3). As we have described previously for the 25R series,^[8,9] treatment of 26-hydroxycholesterol with Jones reagent leads by concomitant allylic oxidation to the undesired 3,6-diketocholest-4-en-26-oic acid. Therefore, we decided to achieve a sequential oxidation of the two hydroxy groups at C-3 and C-26. Selective removal of the silvl ether with TBAF in THF under reflux afforded the 3β-alcohol 17. Oppenauer oxidation of the hydroxy group at C-3 occurred with concomitant isomerisation of the double bond and afforded the

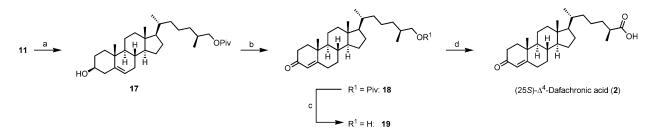
cholest-4-en-3-one 18 in 86% yield. Cleavage of the pivalate proved to be difficult (Table 2). Reaction of the pivalate 18 with bis[tributyltin(IV)] oxide in benzene under reflux resulted in 100% recovery of starting material 18.[21] Saponification of 18 with lithium hydroxide afforded only 29% of the desired 26-hydroxy derivative 19. Using one equivalent of potassium carbonate in methanol at room temperature provided after 6 d the alcohol 19 in 39% yield along with 20% of starting material 18. The best result was obtained by treatment of 18 with sodium methoxide in methanol at room temperature and afforded the alcohol 19 in 56% yield along with 15% of starting material. Completion of the synthesis was achieved by Jones oxidation of the alcohol 19 to provide (25S)- Δ^4 -dafachronic acid (2) in 65% yield.

Table 2. Removal of the pivaloyl protecting group from 18.

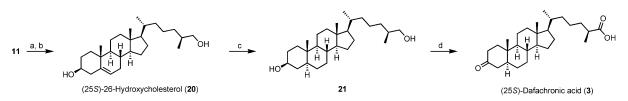
Reaction conditions	% Yield of 19
3.0 equiv. LiOH, MeOH/H ₂ O (10:1), 50 °C, 18 h	29
1.0 equiv. K ₂ CO ₃ , MeOH, 25 °C, 6 d	39 ^[a]
2.0 equiv. NaOMe, MeOH, 25 °C, 5 d	56 ^[b]

[a] 20% of starting material recovered. [b] 15% of starting material recovered.

For the synthesis of (25S)-dafachronic acid (3), both protecting groups had to be removed from our crucial intermediate 11 (Scheme 4). Removal of the pivalate using lithium aluminium hydride and subsequent desilvlation by treatment with TBAF afforded the known (25S)-26-hydroxycho-

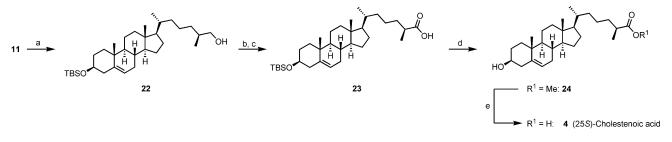


Scheme 3. Synthesis of (25S)-Δ⁴-dafachronic acid (2). Reagents and conditions: (a) 1.5 equiv. TBAF, THF, reflux, 17 h, 92%; (b) 1.5 equiv. Al(O/Pr)₃, acetone/toluene (1:9), 100 °C, 5 h, 86%; (c) 2.0 equiv. NaOMe, MeOH, 25 °C, 5 d, 56% of **19**, 15% of **18**; (d) 5.0 equiv. Jones reagent, 0 °C, 90 min, 65%.



Scheme 4. Synthesis of (25*S*)-dafachronic acid (3). *Reagents and conditions:* (a) 1.5 equiv. TBAF, THF, reflux, 17 h; (b) 4.0 equiv. LiAlH₄, THF, 25 °C, 16 h, 93% for 2 steps; (c) 10% Pd/C, H₂, MeOH/CH₂Cl₂ (1:1), 25 °C, 24 h, 99%; (d) 5.0 equiv. Jones reagent, acetone, 0 °C, 60 min, 88%.

lesterol (20) in only 69% yield.^[22] However, as described above for the synthesis of the diol 16, a reversal of the sequence by first removing the silyl and then the pivaloyl protecting group gave a much better result and provided (25*S*)-26-hydroxycholesterol (20) in 93% yield over both steps. Palladium-catalyzed hydrogenation of 20 led to (25*S*)-5 α cholestan-3 β ,26-diol (21). Finally, Jones oxidation of 21 afforded (25*S*)-dafachronic acid (3) in 88% yield.^[10,11] For the conversion of compound 11 to (25S)-cholestenoic acid (4), the pivaloyl protecting group had to be removed selectively. Treatment of 11 with lithium aluminium hydride afforded the alcohol 22 in 91% yield (Scheme 5). Swern oxidation of 22 followed by oxidation of the crude aldehyde with sodium chlorite provided the silyl-protected (25S)-cholestenoic acid 23 in 89% yield over two steps. In our studies of the 25*R*-series, we found that chromato-



Scheme 5. Transformation of **11** into (25S)-cholestenoic acid (**4**). *Reagents and conditions:* (a) 4.0 equiv. LiAlH₄, THF, 25 °C, 17 h, 91%; (b) 4.0 equiv. DMSO, 2.0 equiv. (COCl)₂, CH₂Cl₂, -78 °C, then **22**, 20 min, 5.0 equiv. Et₃N, to 25 °C; (c) 2.0 equiv. NaClO₂, 10.0 equiv. 2-methyl-2-butene, KH₂PO₄, THF/H₂O (3:1), 25 °C, 24 h, 89% for 2 steps; (d) cat. H₂SO₄, MeOH, reflux, 16 h, 87%; (e) 3.0 equiv. LiOH, THF/MeOH/H₂O (1:1:1), 25 °C, 24 h, 99%.

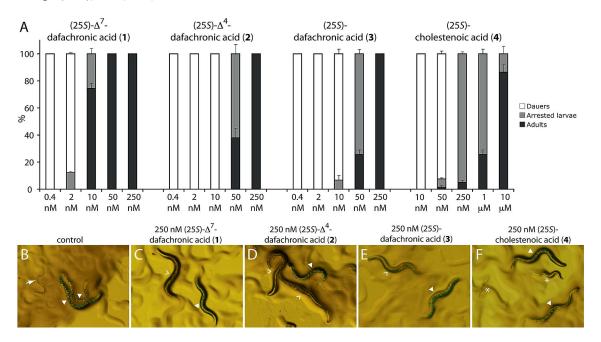


Figure 3. Bioactivity of (25*S*)-dafachronic acids 1–3 and (25*S*)-cholestenoic acid (4). A: rescue of diapause in daf-9(dh6) mutant worms by feeding with the indicated (25*S*)-cholesten-26-oic acids. B: without addition of (25*S*)-cholesten-26-oic acids, daf-9(dh6) mutant worms (no fluorescence) arrest as dauer like larvae (white arrow). C–E: 250 nM (25*S*)-dafachronic acids 1–3 rescue daf-9(dh6) mutant worms to adults (Λ). F: 250 nM (25*S*)-cholestenoic acid (4) rescues only partially daf-9(dh6) mutant worms forming arrested larvae, often with molting defects (*). White triangles in all images indicate fluorescent daf-9(dh6); dhEx24 mutant worms which develop to adults without requirement of exogenous (25*S*)-dafachronic or (25*S*)-cholestenoic acid.



graphic purification of cholestenoic acid is difficult.^[8,9] Thus, esterification was carried out with concomitant cleavage of the silyl ether using catalytic amounts of concentrated sulfuric acid in methanol under reflux. The methyl ester **24** was purified by flash chromatography on silica gel and isolated in 87% yield. A subsequent saponification of the ester with lithium hydroxide provided almost quantitatively pure (25*S*)-cholestenoic acid (**4**).

The bioactivity of 25*S*-steroidal acids 1–4 was investigated by rescue of *daf-9* mutant worms from dauer arrest (Figure 3). The *daf-9(dh6)* mutant worms lacking DAF-9 protein activity cannot generate dafachronic acids.^[1a,3b] In consequence, these mutant worms arrest as dauer-like larvae (Figure 3, A, B). In the experimental setup, some *daf-9(dh6)* mutant worms are obtained as progeny from the strain *daf-9(dh6);dhEx24*.^[3b] The *daf-9(dh6)* mutant worms are identified by the absence of green fluorescence, which is carried by the extrachromosomal array *dhEx24*. In the parental strain *daf-9(dh6);dhEx24*, the extrachromosomal array *dhEx24* rescues the *daf-9(dh6)* mutation (Figure 3, B).^[3b] Normal reproductive development of *daf-9(dh6)* mutant worms is only possible by exogenous supply of dafachronic acid (Figure 3, A–E).

If the supplied amount of dafachronic acid is not sufficient for complete rescue, arrested larvae, often with molting defects, are observed (Figure 3 A, F). Our experiments show that the rescue of daf-9(dh6) mutant worms from dauer arrest is dependent on the concentration of steroidal acids. The rescue of daf-9(dh6) mutant worms from dauer arrest by feeding with the 25S-steroidal acids 1-4 demonstrates the difference in activity for these ligands (Figure 3, A). Our results emphasize that $(25S)-\Delta^7$ -dafachronic acid (1) is the most active ligand. Moreover, it became obvious that (25S)- Δ^4 -dafachronic acid (2) and (25S)-dafachronic acid (3) have a moderate activity, which is about one order of magnitude lower than the activity of 1. The least active compound in this series is (25S)-cholestenoic acid (4) with an activity about one order of magnitude lower than found for the ligands 2 and 3. We have compared the activities of the (25S)-steroidal acids 1-4 with those of the (25R)dafachronic acids, which have been reported in our previous publication.^[9] As a result, the activity of (25R)- Δ^7 -dafachronic acid is in the same range as observed for $(25S)-\Delta^4$ dafachronic acid (2) and (25S)-dafachronic acid (3). While (25R)- Δ^4 -dafachronic acid has an activity comparable to that of (25S)-cholestenoic acid (4).

Conclusions

We have developed a highly efficient synthetic route to all four of the (25S)-cholesten-26-oic acids 1–4 by using a completely stereoselective Evans aldol reaction as key-step. Thus, compound 8 was obtained as a single stereoisomer even on a multigram scale and our crucial intermediate 11 became available in 8 steps and 66% overall yield. Intermediate 11 has been exploited to prepare all four (25S)-steroidal acids: $(25S)-\Delta^7$ -dafachronic acid (1) (15 steps, 27% overall yield), $(25S)-\Delta^4$ -dafachronic acid (2) (12 steps, 19% overall yield), (25S)-dafachronic acid (3) (12 steps, 53% overall yield) and (25S)-cholestenoic acid (4) (13 steps, 46% overall yield). Our present synthesis of the (25S)-cholesten-26-oic acids 1–4 is clearly superior with respect to overall yields and efficiency as compared to the previous approaches to these compounds.^[5,6,11] The present methodology for the assembly of the side chain can be applied to the synthesis of other (25S)-steroids.

Our efficient access to the (25S)-cholesten-26-oic acids 1-4 also set the stage for detailed biological studies towards their hormonal activity in controlling the life cycle of C. elegans. Our study of the biological activity of (25S)dafachronic acids emphasizes the importance of the 3-hydroxy group and suggests that worms can not efficiently oxidise the 3-hydroxy group of (25S)-cholestenoic acid (4). The fact that (25S)- Δ^4 -dafachronic acid (2) and (25S)-dafachronic acid (3) show a similar activity indicates that either the double bond at C-4 is not important for the activity or, that the worms very efficiently convert 3 into 2. The latter reaction might be carried out by a steroid dehydrogenase.^[23] The comparison of the biological activities of the (25S)steroidal acids described in the present publication with those of the (25R)-dafachronic acids reported in our previous paper^[9] emphasizes that the (25S)-steroidal acids are more active than their corresponding (25R)-diastereoisomers. Moreover, it is shown that (25S)- Δ^7 -dafachronic acid (1) has the highest biological activity among all these steroidal acids. The bioactivity of (25R)- Δ^7 -dafachronic acid (1) is in the range of the bioactivity observed for $(25S)-\Delta^4$ dafachronic acid (2) and (25S)-dafachronic acid (3). Whereas (25R)- Δ^4 -dafachronic acid is almost as active as (25S)-cholestenoic acid (4). Thus, by our biological studies the functional groups of the steroids and their stereochemistry required for efficacious signaling can be concluded.

Experimental Section

General: All reactions were carried out in dry solvents and ovendried glassware under argon atmosphere. Tetrahydrofuran, ethyl acetate, dichloromethane, and diethyl ether were dried in a solvent purification system (MBraun-SPS). Acetone was distilled from phosphorus pentoxide and stored over molecular sieves (3 Å). Benzene was dried with sodium. Toluene was purchased from Acros Organics (water content less than 50 ppm). Dry methanol was purchased from VWR Prolabo (water content less than 20 ppm). Pyridine was obtained from Fluka (water content less than 50 ppm). Triethylamine was heated under reflux with calcium hydride for 48 h and stored over 3 Å molecular sieves. Commercial AIBN was recrystallised from methanol. Dibutylboron triflate was obtained from Acros Organics as a 1 M solution in dichloromethane. All other chemicals were used as received from commercial sources. Flash chromatography was performed using silica gel from Acros Organics (0.063-0.200 mm). Thin layer chromatography was performed with TLC plates from Merck (60 F₂₅₄) using anisaldehyde solution for visualization. Melting points were measured on an Electrothermal IA9100 melting point apparatus. Specific rotation values were obtained from a Perkin-Elmer 341 polarimeter. Infra-

red spectra were recorded on a Thermo Nicolet Avatar 360 FT-IR spectrometer using the ATR method (Attenuated Total Reflectance). NMR spectra were recorded on a Bruker DRX 500 NMR spectrometer. Complete assignment of the ¹H and ¹³C signals was achieved by HSQC experiments. Chemical shifts δ are reported in ppm with the deuterated solvent as internal standard. The following abbreviations have been used: s: singlet, d: doublet, dd: doublet of doublets, dt: doublet of triplets, t: triplet, q: quartet, sext: sextet, sept: septet, m: multiplet, br: broad. Mass spectra were recorded on a Finnigan MAT-95 spectrometer (electron impact, 70 eV) or by GC/MS-coupling using an Agilent Technologies 6890N GC System equipped with a 5973 Mass Selective Detector (electron impact EI, 70 eV). ESI-MS spectra were recorded on an Esquire LC with an ion trap detector from Bruker. Positive and negative ions were detected. Elemental analyses were measured on an EuroVector EuroEA3000 elemental analyser. X-ray analyses: Bruker-Nonius Kappa CCD equipped with a 700 series Cryostream low temperature device from Oxford Cryosystems and STOE IPDS 2 image plate. Software: Collect (Nonius BV, 1999), Dirax/lsg (Duisenberg, 1992), SHELXS-97 (G. M. Sheldrick, 1990), EvalCCD (Duisenberg et al., 2003), SADABS version 2.10 (G. M. Sheldrick, Bruker AXS Inc., 2002), SHELXL-97 (G. M. Sheldrick, 1997), Schakal-99 (E. Keller, 1999).

3β-(tert-Butyldimethylsilyloxy)chol-5-en-24-ol (6): tert-Butylchloro-(3.78 g, 25.09 mmol), (4.32 g, dimethylsilane imidazole 63.50 mmol) and DMAP (2.2 g, 17.25 mmol) were added to a solution of 3β-hydroxychol-5-en-24-oic acid (5) (2.936 g, 7.84 mmol) in DMF (50 mL). Additional DMF (50 mL) was added and the solution was stirred at room temperature for 18 h. After addition of water (250 mL), the resulting mixture was extracted with diethyl ether $(3 \times 50 \text{ mL})$. The combined organic layers were washed with water $(2 \times 100 \text{ mL})$, brine (100 mL) and then dried with magnesium sulfate. Evaporation of the solvent gave the crude product which was dissolved in THF (50 mL). Lithium aluminium hydride (1.19 g, 31.36 mmol) was added in portions to this solution at 0 °C and the resulting mixture was stirred at room temperature for 17 h. Water (50 mL) was slowly added and the mixture was extracted with diethyl ether $(3 \times 50 \text{ mL})$. The combined organic layers were dried with magnesium sulfate and the solvent was removed. Purification of the residue by flash chromatography on silica gel (petroleum ether/diethyl ether, 4:1) provided the alcohol 6, yield 3.561 g (96%). Colourless solid; m.p. 171-173 °C (ref.[15] 146.5-152 °C). ¹H NMR (500 MHz, CDCl₃): δ = 0.04 (s, 6 H), 0.67 (s, 3 H), 0.88 (s, 9 H), 0.93 (d, J = 6.6 Hz, 3 H), 0.95–1.17 (m, 6 H), 0.98 (s, 3 H), 1.21– 1.27 (m, 2 H), 1.39–1.72 (m, 11 H), 1.79 (dt, J = 13.3, 3.5 Hz, 1 H), 1.82-1.84 (m, 1 H), 1.93-2.01 (m, 2 H), 2.15 (ddd, J = 13.3, 4.9, 2.2 Hz, 1 H), 2.25 (m, 1 H), 3.47 (m, 1 H), 3.60 (m, 2 H), 5.30 (m, 1 H) ppm. ¹³C NMR and DEPT (125 MHz, CDCl₃): $\delta = -4.60$ (2 CH₃), 11.84 (CH₃), 18.27 (C), 18.66 (CH₃), 19.42 (CH₃), 21.03 (CH₂), 24.25 (CH₂), 25.93 (3 CH₃), 28.22 (CH₂), 29.37 (CH₂), 31.82 (CH₂), 31.87 (CH), 31.90 (CH₂), 32.06 (CH₂), 35.55 (CH), 36.56 (C), 37.36 (CH₂), 39.76 (CH₂), 42.31 (C), 42.79 (CH₂), 50.15 (CH), 55.95 (CH), 56.76 (CH), 63.60 (CH₂), 72.63 (CH), 121.13 (CH), 141.54 (C) ppm. C₃₀H₅₄O₂Si (474.83): C 75.88, H 11.46, found: C 75.94, H 11.34%. For further spectroscopic data see ref.^[15]

3*β*-(*tert*-**Butyldimethylsilyloxy)chol-5-en-24-al (7):** Oxalyl chloride (396 μ L, 4.68 mmol) was added slowly to a solution of DMSO (665 μ L, 9.36 mmol) in dichloromethane (10 mL) at -78 °C. After 5 min, a solution of the alcohol **6** (1.109 g, 2.34 mmol) in dichloromethane (20 mL) was added and stirring was continued at -78 °C for 20 min. Then, triethylamine (1.63 mL, 11.70 mmol) was added, the solution was warmed to room temperature and stirring was continued for additional 10 min. The reaction mixture was

quenched by addition of a saturated aqueous solution of ammonium chloride (20 mL). The layers were separated and the aqueous layer was extracted with dichloromethane $(2 \times 50 \text{ mL})$. The combined organic layers were dried with magnesium sulfate and the solvent was evaporated. Purification of the residue by flash chromatography on silica gel (petroleum ether/diethyl ether, 10:1) afforded the aldehyde 7, yield 1.081 g (98%). Colourless solid; m.p. 137–139 °C (ref.^[15] 132–137 °C). ¹H NMR (500 MHz, CDCl₃): δ = 0.04 (s, 6 H), 0.66 (s, 3 H), 0.87 (s, 9 H), 0.91 (d, J = 6.5 Hz, 3 H), 0.95-1.18 (m, 6 H), 0.98 (s, 3 H), 1.24-1.34 (m, 3 H), 1.40-1.60 (m, 6 H), 1.68-1.86 (m, 4 H), 1.93-2.00 (m, 2 H), 2.15 (ddd, J = 13.3, 4.9, 2.2 Hz, 1 H), 2.25 (m, 1 H), 2.35 (m, 1 H), 2.44 (m, 1 H), 3.47 (m, 1 H), 5.30 (m, 1 H), 9.76 (t, J = 1.9 Hz, 1 H) ppm. ¹³C NMR and DEPT (125 MHz, CDCl₃): $\delta = -4.60$ (2 CH₃), 11.85 (CH₃), 18.26 (C), 18.40 (CH₃), 19.41 (CH₃), 21.02 (CH₂), 24.23 (CH₂), 25.93 (3 CH₃), 27.94 (CH₂), 28.17 (CH₂), 31.87 (CH, CH₂), 32.05 (CH₂), 35.32 (CH), 36.55 (C), 37.35 (CH₂), 39.72 (CH₂), 40.92 (CH₂), 42.37 (C), 42.78 (CH₂), 50.11 (CH), 55.76 (CH), 56.73 (CH), 72.60 (CH), 121.09 (CH), 141.54 (C), 203.25 (CHO) ppm. C₃₀H₅₂O₂Si (472.82): C 76.21, H 11.09, found: C 76.27, H 11.14%. For further spectroscopic data see ref.^{[15].}

 $(4S,\!24'R,\!25'S)\!-\!3\!-\![3'\beta-(tert-Butyldimethylsilyloxy)\!-\!24'-hydroxychol$ est-5'-en-26'-oyl]-4-isopropyloxazolidin-2-one (8): A 1.0 M solution of dibutylboron triflate (3.28 mL, 3.28 mmol) was added to a solution of (S)-(+)-4-isopropyl-3-propionyl-2-oxazolidinone (505 µL, 2.98 mmol) in dichloromethane (10 mL) at 0 °C. After 5 min, triethylamine (539 µL, 3.87 mmol) was added dropwise and the resulting mixture was stirred at 0 °C for 1 h. The reaction mixture was cooled to -78 °C and a solution of the aldehyde 7 (1.081 g, 2.29 mmol) in dichloromethane (10 mL) was added dropwise. Stirring was continued at -78 °C for 30 min. Then, the mixture was warmed to 0 °C and stirring was continued for additional 80 min. Methanol (10 mL) and 30% aqueous H₂O₂ (10 mL) were added and the resulting mixture was stirred at 0 °C for 30 min. Water (100 mL) was added and the layers were separated. The aqueous layer was extracted with dichloromethane $(2 \times 50 \text{ mL})$. The combined organic layers were dried with magnesium sulfate and the solvent was evaporated. Purification of the residue by flash chromatography on silica gel (petroleum ether/diethyl ether, 4:1 to 2:1) afforded the aldol product 8 as a single stereoisomer, yield 1.428 g (95%). Colourless solid; m.p. 172–174 °C. IR (ATR): v = 3533, 2928, 2858, 1773, 1701, 1680, 1458, 1386, 1367, 1302, 1236, 1207, 1143, 1078, 1057, 1017, 959, 886, 870, 835, 807, 774, 719 cm^{-1} . ¹H NMR (500 MHz, CDCl₃): $\delta = 0.04$ (s, 6 H), 0.66 (s, 3 H), 0.87 (d, J = 6.5 Hz, 3 H), 0.87 (s, 9 H), 0.89-1.28 (m, 9 H), 0.91(d, J = 7.1 Hz, 6 H), 0.98 (s, 3 H), 1.24 (d, J = 7.1 Hz, 3 H), 1.38– 1.60 (m, 9 H), 1.70 (m, 1 H), 1.79 (dt, J = 13.3, 3.3 Hz, 1 H), 1.82-1.85 (m, 1 H), 1.93–2.00 (m, 2 H), 2.15 (ddd, J = 13.3, 4.8, 2.1 Hz, 1 H), 2.25 (m, 1 H), 2.33 (dsept, J = 4.0, 7.0 Hz, 1 H), 2.91 (br. d, J = 2.1 Hz, 1 H), 3.46 (m, 1 H), 3.76 (dq, J = 2.6, 7.1 Hz, 1 H), 3.87 (m, 1 H), 4.21 (dd, J = 8.6, 3.3 Hz, 1 H), 4.27 (t, J = 8.6 Hz, 1 H), 4.46 (ddd, J = 8.6, 4.0, 3.3 Hz, 1 H), 5.30 (m, 1 H) ppm. ¹³C NMR and DEPT (125 MHz, CDCl₃): $\delta = -4.61$ (2 CH₃), 10.78 (CH₃), 11.84 (CH₃), 14.66 (CH₃), 17.90 (CH₃), 18.25 (C), 18.66 (CH₃), 19.41 (CH₃), 21.02 (CH₂), 24.25 (CH₂), 25.92 (3 CH₃), 28.18 (CH₂), 28.30 (CH), 30.07 (CH₂), 31.87 (CH), 31.89 (CH₂), 32.03 (CH₂), 32.06 (CH₂), 35.45 (CH), 36.55 (C), 37.34 (CH₂), 39.73 (CH₂), 42.08 (CH), 42.30 (C), 42.79 (CH₂), 50.13 (CH), 55.79 (CH), 56.73 (CH), 58.18 (CH), 63.29 (CH₂), 71.54 (CH), 72.61 (CH), 121.14 (CH), 141.52 (C), 153.50 (C=O), 177.93 (C=O) ppm. ESI-MS: $m/z = 658.5 [(M + H)^+], 640.4, 508.4. C_{39}H_{67}NO_5Si$ (658.04): C 71.18, H 10.26, N 2.13, found: C 71.43, H 10.34, N 2.25%.



(24R,25R)-3β-(tert-Butyldimethylsilyloxy)-26-(pivaloyloxy)cholest-5en-24-ol (9): A 2.0 M solution of lithium borohydride in THF (908 μ L, 1.815 mmol) and water (30 μ L, 1.65 mmol) were added to a solution of the aldol product 8 (1.085 g, 1.65 mmol) in diethyl ether (25 mL) at 0 °C. After stirring for 2 h at 0 °C, water (20 mL) and 2 N NaOH (20 mL) were slowly added to the reaction mixture, which was subsequently extracted with dichloromethane $(2 \times 50 \text{ mL})$, diethyl ether $(2 \times 50 \text{ mL})$ and ethyl acetate $(2 \times 50 \text{ mL})$. The combined organic layers were dried with magnesium sulfate and the solvent was evaporated. The crude diol was dissolved in pyridine (15 mL) and dichloromethane (15 mL). After addition of pivaloyl chloride (264 µL, 2.145 mmol) at 0 °C, the resulting mixture was stirred at 0 °C for 2 h and then water (50 mL) was added. The mixture was extracted with dichloromethane $(3 \times 50 \text{ mL})$ and the combined organic layers were dried with magnesium sulfate. Evaporation of the solvent and purification of the residue by flash chromatography on silica gel (petroleum ether/diethyl ether, 6:1) provided the pivalate 9, yield 818 mg (81%). Colourless solid; m.p. 133–136 °C. IR (ATR): $\tilde{v} = 3525$, 2958, 2926, 2898, 2851, 1710, 1471, 1459, 1382, 1287, 1247, 1174, 1131, 1076, 1029, 1010, 975, 940, 892, 873, 834, 805, 777 cm^{-1} . ¹H NMR (500 MHz, CDCl₃): $\delta = 0.04$ (s, 6 H), 0.66 (s, 3 H), 0.88 (s, 9 H), 0.90–1.33 (m, 8 H), 0.911 (d, J = 7.0 Hz, 3 H), 0.915 (d, *J* = 6.4 Hz, 3 H), 0.98 (s, 3 H), 1.20 (s, 9 H), 1.37–1.58 (m, 10 H), 1.69–1.74 (m, 1 H), 1.79 (dt, J = 13.3, 3.3 Hz, 1 H), 1.82–1.87 (m, 2 H), 1.94 (m, 1 H), 1.99 (dt, J = 12.4, 3.3 Hz, 1 H), 2.15 (ddd, J = 13.3, 4.9, 2.2 Hz, 1 H), 2.25 (m, 1 H), 3.47 (m, 1 H), 3.53 (m, 1 H), 3.92 (dd, J = 11.0, 5.8 Hz, 1 H), 4.19 (dd, J = 11.0, 7.6 Hz, 1 H)1 H), 5.30 (m, 1 H) ppm. ¹³C NMR and DEPT (125 MHz, $CDCl_3$): $\delta = -4.60 (2 CH_3), 10.27 (CH_3), 11.84 (CH_3), 18.26 (C),$ 18.67 (CH₃), 19.41 (CH₃), 21.03 (CH₂), 24.25 (CH₂), 25.93 (3 CH₃), 27.21 (3 CH₃), 28.26 (CH₂), 30.60 (CH₂), 31.87 (CH), 31.90 (CH₂), 32.06 (CH₂), 32.28 (CH₂), 35.50 (CH), 36.56 (C), 37.36 (CH₂), 37.95 (CH), 38.83 (C), 39.75 (CH₂), 42.31 (C), 42.79 (CH₂), 50.14 (CH), 55.83 (CH), 56.75 (CH), 66.83 (CH₂), 71.88 (CH), 72.62 (CH), 121.13 (CH), 141.54 (C), 178.89 (C=O) ppm. GC-MS (70 eV): m/z (%) = 559 (29) [(M - tBu)⁺], 383 (18), 365 (16), 341 (27), 331 (11), 255 (15), 211 (20), 175 (11), 171 (11), 161 (33), 159 (100). C₃₈H₆₈O₄Si (617.03): C 73.97, H 11.11, found: C 74.04, H 11.19%.

O-(24R,25R)-3\beta-(tert-Butyldimethylsilyloxy)-26-(pivaloyloxy)cholest-5-en-24-yl S-Methyl Xanthate (10): A 1.0 M solution of NaHMDS in THF (136 µL, 136 µmol) and carbon disulfide (164 μ L, 2.72 mmol) were added to a solution of the alcohol 9 (84 mg, 136 µmol) in THF at -78 °C. After stirring for 30 min at -78 °C, the solution was warmed to 0 °C and iodomethane (17 μ L, 272 µmol) was added. Stirring was continued for additional 30 min, water (50 mL) was added and the solution was extracted with diethyl ether $(3 \times 50 \text{ mL})$. The combined organic layers were dried with magnesium sulfate and the solvent was removed to provide the xanthate 10, yield 94 mg (98%). Yellow oil (which crystallised on standing). ¹H NMR (500 MHz, CDCl₃): $\delta = 0.04$ (s, 6 H), 0.65 (s, 3 H), 0.85-0.93 (m, 1 H), 0.88 (s, 9 H), 0.91 (d, J = 6.5 Hz, 3 H), 0.95-1.26 (m, 7 H), 0.98 (s, 3 H), 1.02 (d, J = 7.0 Hz, 3 H), 1.20 (s, 9 H), 1.36-1.57 (m, 8 H), 1.70 (m, 1 H), 1.77-1.83 (m, 3 H), 1.93-1.98 (m, 2 H), 2.12-2.27 (m, 4 H), 2.54 (s, 3 H), 3.47 (m, 1 H), 3.97 (d, J = 6.5 Hz, 2 H), 5.30 (m, 1 H), 5.78 (m, 1 H) ppm. ¹³C NMR and DEPT (125 MHz, CDCl₃): δ = -4.60 (2 CH₃), 11.85 (2 CH₃), 18.27 (C), 18.57 (CH₃), 18.87 (CH₃), 19.41 (CH₃), 21.03 (CH₂), 24.23 (CH₂), 25.93 (3 CH₃), 27.05 (CH₂), 27.20 (3 CH₃), 28.02 (CH₂), 31.31 (CH₂), 31.88 (CH, CH₂), 32.06 (CH₂), 35.17 (CH), 36.08 (CH), 36.56 (C), 37.36 (CH₂), 38.78 (C), 39.74 (CH₂), 42.31 (C), 42.79 (CH₂), 50.14 (CH), 55.49 (CH), 56.73 (CH), 65.56

(CH₂), 72.61 (CH), 84.56 (CH), 121.12 (CH), 141.55 (C), 178.41 (C=O), 215.94 (C=S) ppm.

(25S)-3β-(tert-Butyldimethylsilyloxy)-26-(pivaloyloxy)cholest-5-ene (11): AIBN (3.5 mg, 21.1 µmol) was added to a solution of the xanthate 10 (149 mg, 211 µmol) in toluene (5 mL). The flask containing this mixture was put in an oil bath, which was already heated at 110 °C. A solution of tributylstannane (851 µL, 3.165 mmol) in toluene (2 mL) was slowly added to the hot reaction mixture. After 5 min, the yellow solution became colourless, indicating complete conversion. After cooling to room temperature, a saturated aqueous solution of sodium hydrogen carbonate (50 mL) was added and the mixture was extracted with diethyl ether $(3 \times 50 \text{ mL})$. The combined organic layers were dried with magnesium sulfate and the solvent was evaporated. Purification of the residue by flash chromatography on silica gel (petroleum ether/diethyl ether, 100:1 to 70:1) afforded compound 11, yield 118 mg (93%). Colourless solid; m.p. 116–118 °C. IR (ATR): $\tilde{v} = 2929$, 2895, 2856, 1728, 1471, 1397, 1368, 1282, 1252, 1152, 1090, 1033, 989, 959, 939, 890, 870, 835, 804, 771, 666 cm⁻¹. ¹H NMR $(500 \text{ MHz}, \text{CDCl}_3)$: $\delta = 0.04$ (s, 6 H), 0.66 (s, 3 H), 0.86–1.28 (m, 10 H), 0.88 (s, 9 H), 0.90 (d, J = 6.6 Hz, 3 H), 0.92 (d, J = 6.8 Hz, 3 H), 0.98 (s, 3 H), 1.19 (s, 9 H), 1.30-1.59 (m, 10 H), 1.69-1.83 (m, 4 H), 1.94 (m, 1 H), 1.99 (dt, J = 13.0, 3.3 Hz, 1 H), 2.15 (ddd, J = 13.3, 4.9, 2.1 Hz, 1 H), 2.26 (m, 1 H), 3.47 (m, 1 H), 3.82 (dd, J = 10.7, 6.7 Hz, 1 H), 3.93 (dd, J = 10.7, 5.6 Hz, 1 H), 5.30 (m, 1 H) ppm. ¹³C NMR and DEPT (125 MHz, CDCl₃): $\delta = -4.60$ (2 CH₃), 11.82 (CH₃), 17.07 (CH₃), 18.27 (C), 18.70 (CH₃), 19.42 (CH₃), 21.04 (CH₂), 23.20 (CH₂), 24.27 (CH₂), 25.93 (3 CH₃), 27.22 (3 CH₃), 28.22 (CH₂), 31.88 (CH), 31.92 (CH₂), 32.07 (CH₂), 32.70 (CH), 33.86 (CH₂), 35.67 (CH), 36.14 (CH₂), 36.57 (C), 37.36 (CH₂), 38.84 (C), 39.77 (CH₂), 42.30 (C), 42.80 (CH₂), 50.17 (CH), 56.01 (CH), 56.77 (CH), 69.11 (CH₂), 72.63 (CH), 121.14 (CH), 141.56 (C), 178.64 (C=O) ppm. MS (70 eV): m/z (%) = 600 (1) $[M^+]$, 585 (3), 543 (73) $[(M - tBu)^+]$, 441 (12), 367 (100), 295 (11), 255 (14), 161 (20), 159 (79). HRMS: m/z calcd. for C₃₄H₅₉O₃Si $[(M - tBu)^+]$: 543.4233, found: 543.4246.

(25S)-3β-(tert-Butyldimethylsilyloxy)-26-(pivaloyloxy)cholest-5-en-7-one (12): A 5.5 M solution of tert-butyl hydroperoxide in decane (629 µL, 3.46 mmol), PDC (1.30 g, 3.46 mmol) and Celite[®] (1.03 g) were added to a solution of compound 11 (520 mg, 0.865 mmol) in benzene (30 mL) at 0 °C. The resulting mixture was stirred at room temperature for 24 h. A second portion of tert-butyl hydroperoxide (629 μ L, 3.46 mmol) was added and stirring was continued for 17 h. The reaction mixture was filtered through a short pad of silica gel with diethyl ether (500 mL) and the solvent was removed. Purification of the residue by flash chromatography on silica gel (petroleum ether/diethyl ether, 20:1) provided the enone 12, yield 398 mg (75%). Colourless crystals; m.p. 153–155 °C. IR (ATR): $\tilde{v} = 2934$, 2858, 1730, 1666, 1626, 1461, 1375, 1282, 1253, 1153, 1091, 1033, 955, 937, 892, 877, 835, 804, 771 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): $\delta = 0.05$ (s, 6 H), 0.66 (s, 3 H), 0.88 (s, 9 H), 0.90 (d, J =6.4 Hz, 3 H), 0.91 (d, J = 6.7 Hz, 3 H), 1.00–1.66 (m, 17 H), 1.17 (s, 3 H), 1.19 (s, 9 H), 1.74–1.82 (m, 2 H), 1.85–1.91 (m, 2 H), 2.01 (dt, J = 12.7, 3.3 Hz, 1 H), 2.22 (dd, J = 12.3, 10.9 Hz, 1 H), 2.33-2.43 (m, 3 H), 3.59 (m, 1 H), 3.83 (dd, J = 10.7, 6.7 Hz, 1 H), 3.93 (dd, J = 10.7, 5.6 Hz, 1 H), 5.65 (d, J = 1.4 Hz, 1 H) ppm. ¹³C NMR and DEPT (125 MHz, CDCl₃): $\delta = -4.70$ (CH₃), -4.67(CH₃), 11.93 (CH₃), 17.07 (CH₃), 17.28 (CH₃), 18.15 (C), 18.85 (CH₃), 21.16 (CH₂), 23.26 (CH₂), 25.82 (3 CH₃), 26.28 (CH₂), 27.22 (3 CH₃), 28.54 (CH₂), 31.71 (CH₂), 32.70 (CH), 33.83 (CH₂), 35.61 (CH), 36.15 (CH₂), 36.39 (CH₂), 38.34 (C), 38.68 (CH₂), 38.85 (C), 42.51 (CH₂), 43.05 (C), 45.38 (CH), 49.90 (CH), 49.95 (CH), 54.66 (CH), 69.10 (CH₂), 71.31 (CH), 125.78 (CH), 165.89 (C), 178.65

(C=O), 202.47 (C=O) ppm. GC-MS (70 eV): m/z (%) = 614 (2) [M⁺], 557 (41), 381 (100), 269 (25). C₃₈H₆₆O₄Si (615.01): C 74.21, H 10.82, found: C 74.24, H 10.92%.

(25S)-3β-(tert-Butyldimethylsilyloxy)-26-(pivaloyloxy)cholest-5α-an-7-one (13): A solution of the enone 12 (100 mg, 163 μ mol) in ethyl acetate (10 mL) was added to a Schlenk flask, loaded with Pd/C (10%, 17.3 mg, 16.3 µmol Pd). The resulting mixture was stirred at room temperature for 16 h under an hydrogen atmosphere. Filtration of the mixture over a short pad of Celite® with ethyl acetate, evaporation of the solvent and purification of the crude product by flash chromatography on silica gel (petroleum ether/diethyl ether, 10:1) afforded the ketone 13, yield 95 mg (95%). Colourless solid; m.p. 116-117 °C. IR (ATR): v = 2929, 2857, 1726, 1704, 1471, 1375, 1284, 1250, 1162, 1099, 1054, 1007, 986, 943, 873, 835, 797, 772 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): $\delta = 0.025$ (s, 3 H), 0.026 (s, 3 H), 0.63 (s, 3 H), 0.86 (s, 9 H), 0.89 (d, J = 6.7 Hz, 3 H), 0.91 (d, J = 6.8 Hz, 3 H), 0.93–1.56 (m, 20 H), 1.06 (s, 3 H), 1.19 (s, 9 H), 1.69–1.78 (m, 3 H), 1.85–1.89 (m, 1 H), 1.94–2.00 (m, 2 H), 2.16 (m, 1 H), 2.33 (m, 2 H), 3.53 (m, 1 H), 3.82 (dd, J =10.7, 6.7 Hz, 1 H), 3.92 (dd, J = 10.7, 5.6 Hz, 1 H) ppm. ¹³C NMR and DEPT (125 MHz, CDCl₃): $\delta = -4.66$ (2 CH₃), 11.85 (CH₃), 12.02 (CH₃), 17.05 (CH₃), 18.20 (C), 18.75 (CH₃), 21.83 (CH₂), 23.23 (CH₂), 24.92 (CH₂), 25.87 (3 CH₃), 27.21 (3 CH₃), 28.40 (CH₂), 31.52 (CH₂), 32.71 (CH), 33.83 (CH₂), 35.55 (CH), 36.01 (C), 36.12 (CH₂), 36.21 (CH₂), 38.44 (CH₂), 38.74 (CH₂), 38.84 (C), 42.47 (C), 46.20 (CH₂), 47.13 (CH), 48.83 (CH), 49.99 (CH), 54.92 (CH), 55.44 (CH), 69.10 (CH₂), 71.50 (CH), 178.64 (C=O), 212.44 (C=O) ppm. GC-MS (70 eV): m/z (%) = 601 (3) [(M -Me)⁺], 561 (8), 559 (59), 457 (28), 383 (29), 365 (25), 271 (31), 253 (11), 177 (11), 161 (35), 159 (89), 75 (100), 57 (73). C₃₈H₆₈O₄Si (617.03): C 73.97, H 11.11, found: C 74.06, H 11.22%.

(25S)-3β-(tert-Butyldimethylsilyloxy)-26-(pivaloyloxy)cholest-5α-an-7α-ol (14): A 1.0 M solution of L-Selectride[®] in THF (754 µL, 754 µmol) was added to a solution of the ketone 13 (358 mg, 580 µmol) in THF (20 mL) at -78 °C. Stirring was continued at the same temperature for 1.5 h. The reaction mixture was quenched by addition of a saturated aqueous solution of sodium hydrogen carbonate (10 mL), methanol (5 mL) and 30% aqueous H_2O_2 (5 mL), and was warmed to 0 °C. After stirring for 30 min, water (50 mL) and diethyl ether (50 mL) were added and the layers were separated. The aqueous layer was extracted with diethyl ether $(2 \times 50 \text{ mL})$ and the combined organic layers were dried with magnesium sulfate. Evaporation of the solvent and purification of the residue by flash chromatography on silica gel (petroleum ether/diethyl ether, 10:1) provided the 7 α -alcohol 14 as a single stereoisomer, yield 324 mg (90%). Colourless solid; m.p. 90-92 °C. IR (ATR): $\tilde{v} = 3501, 2929, 2855, 1728, 1708, 1471, 1398, 1380, 1285,$ 1250, 1162, 1101, 1075, 1032, 1006, 975, 947, 871, 835, 814, 772 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): $\delta = 0.03$ (s, 6 H), 0.63 (s, 3 H), 0.78 (s, 3 H), 0.86 (s, 9 H), 0.89 (d, J = 6.5 Hz, 3 H), 0.91 (d, J = 6.8 Hz, 3 H), 0.98–1.05 (m, 2 H), 1.06–1.16 (m, 6 H), 1.19 (s, 9 H), $1.21-1.68 \text{ (m, 17 H)}, 1.56 \text{ (dt, } J = 12.6, 3.1 \text{ Hz}, 1 \text{ H)}, 1.74-1.86 \text{ (m, 17 H)$ 2 H), 1.92 (dt, J = 12.6, 3.2 Hz, 1 H), 3.57 (m, 1 H), 3.81 (m, 1 H), 3.82 (dd, J = 10.7, 6.7 Hz, 1 H), 3.93 (dd, J = 10.7, 5.6 Hz, 1 H)ppm. ¹³C NMR and DEPT (125 MHz, CDCl₃): δ = -4.60 (CH₃), -4.58 (CH₃), 11.26 (CH₃), 11.80 (CH₃), 17.05 (CH₃), 18.25 (C), 18.63 (CH₃), 20.95 (CH₂), 23.17 (CH₂), 23.63 (CH₂), 25.94 (3 CH₃), 27.21 (3 CH₃), 28.19 (CH₂), 31.85 (CH₂), 32.71 (CH), 33.85 (CH₂), 35.57 (C), 35.66 (CH), 36.10 (CH₂), 36.29 (CH₂), 36.90 (CH₂), 37.20 (CH), 38.17 (CH₂), 38.84 (C), 39.50 (CH₂), 39.53 (CH), 42.65 (C), 45.91 (CH), 50.57 (CH), 55.99 (CH), 68.11 (CH), 69.11 (CH₂), 71.96 (CH), 178.64 (C=O) ppm. GC-MS (70 eV): m/z (%) = 603 (2) $[(M - Me)^+]$, 561 (13), 485 (18), 451 (12), 383 (13), 378 (15),

367 (17), 345 (14), 343 (10), 331 (76), 281 (24), 273 (16), 269 (32), 259 (14), 257 (22), 255 (13), 213 (14), 211 (23), 75 (100). $C_{38}H_{70}O_4Si$ (619.05): C 73.73, H 11.40, found: C 73.84, H 11.48%.

(25S)-3β-(tert-Butyldimethylsilyloxy)-26-(pivaloyloxy)-5α-cholest-7ene (15): Thionyl chloride (41 µL, 565 µmol) was added to a solution of the 7 α -alcohol 14 (70 mg, 113 μ mol) in pyridine (5 mL) at 0 °C. The resulting mixture was stirred at 0 °C for 40 min and the solvent was evaporated. Purification of the residue by flash chromatography on silica gel (petroleum ether/diethyl ether, 50:1) afforded the cholest-7-ene 15, yield 59 mg (87%). Colourless solid; m.p. 79–80 °C. IR (ATR): \tilde{v} = 2930, 2854, 1728, 1471, 1398, 1377, 1282, 1252, 1161, 1102, 1084, 1006, 982, 940, 871, 835, 815, 795, 772 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): $\delta = 0.04$ (s, 6 H), 0.51 (s, 3 H), 0.77 (s, 3 H), 0.87 (s, 9 H), 0.90 (d, J = 6.4 Hz, 3 H), 0.92 (d, J = 6.7 Hz, 3 H), 0.99–1.80 (m, 26 H), 1.19 (s, 9 H), 1.83–1.87 (m, 1 H), 2.00 (dt, J = 12.3, 3.2 Hz, 1 H), 3.53 (m, 1 H), 3.83 (dd, J =10.7, 6.8 Hz, 1 H), 3.93 (dd, J = 10.7, 5.6 Hz, 1 H), 5.14 (m, 1 H) ppm. ¹³C NMR and DEPT (125 MHz, CDCl₃): $\delta = -4.58$ (2 CH₃), 11.80 (CH₃), 13.07 (CH₃), 17.06 (CH₃), 18.28 (C), 18.83 (CH₃), 21.49 (CH₂), 22.93 (CH₂), 23.28 (CH₂), 25.94 (3 CH₃), 27.21 (3 CH₃), 27.94 (CH₂), 29.69 (CH₂), 31.86 (CH₂), 32.69 (CH), 33.84 (CH₂), 34.21 (C), 36.08 (CH, CH₂), 37.31 (CH₂), 38.43 (CH₂), 38.84 (C), 39.56 (CH₂), 40.37 (CH), 43.37 (C), 49.50 (CH), 55.02 (CH), 56.02 (CH), 69.11 (CH₂), 71.89 (CH), 117.53 (CH), 139.53 (C), 178.65 (C=O) ppm. GC-MS (70 eV): m/z (%) = 600 (6) [M⁺], 585 (7), 545 (9), 543 (74), 468 (21), 467 (20), 441 (22), 367 (15), 255 (10), 161 (19), 159 (69), 75 (100). C₃₈H₆₈O₃Si (601.03): C 75.94, H 11.40, found: C 75.82, H 11.40%.

(25S)-26-Hydroxy-5α-cholest-7-en-3β-ol (16): A 1.0 M solution of tetrabutylammonium fluoride in THF (0.53 mL, 530 µmol) was added to a solution of the cholest-7-ene 15 (214 mg, 356 µmol) in THF (15 mL). The mixture was heated under reflux for 16 h. After cooling to room temperature, water was added (50 mL) and the resulting mixture was extracted with diethyl ether (3×50 mL). The combined organic layers were dried with magnesium sulfate and the solvent was evaporated. The crude product was dissolved in THF (10 mL) and lithium aluminium hydride (54 mg, 1.424 mmol) was added at 0 °C. The resulting mixture was stirred at room temperature for 17 h. Then, 10% sulfuric acid (10 mL) and water (50 mL) were added. The mixture was extracted with ethyl acetate $(3 \times 50 \text{ mL})$ and the combined organic layers were dried with magnesium sulfate. Evaporation of the solvent and purification of the residue by flash chromatography on silica gel (petroleum ether/ ethyl acetate, 3:1) provided the diol 16, which has been recrystallised from ethyl acetate, yield 118 mg (82%). Colourless crystals; m.p. 165–167 °C. IR (ATR): $\tilde{v} = 3303$, 2914, 2865, 1464, 1447, 1381, 1366, 1347, 1100, 1052, 1032, 1019, 976, 941, 848, 832, 729 cm^{-1} . ¹H NMR (500 MHz, CDCl₃): $\delta = 0.52$ (s, 3 H), 0.78 (s, 3 H), 0.91 (d, J = 6.7 Hz, 6 H), 0.98-1.30 (m, 9 H), 1.33-1.64 (m, 12 H),1.69-1.90 (m, 6 H), 2.01 (dt, J = 12.7, 3.4 Hz, 1 H), 3.41 (dd, J = 12.7, 3.4 Hz, 1 H),10.5, 6.5 Hz, 1 H), 3.50 (dd, J = 10.5, 5.7 Hz, 1 H), 3.58 (m, 1 H), 5.14 (dd, J = 4.7, 2.2 Hz, 1 H) ppm. ¹³C NMR and DEPT (125 MHz, CDCl₃): δ = 11.84 (CH₃), 13.04 (CH₃), 16.72 (CH₃), 18.85 (CH₃), 21.52 (CH₂), 22.93 (CH₂), 23.51 (CH₂), 27.95 (CH₂), 29.62 (CH₂), 31.45 (CH₂), 33.63 (CH₂), 34.18 (C), 35.81 (CH), 36.17 (CH, CH₂), 37.11 (CH₂), 37.95 (CH₂), 39.53 (CH₂), 40.21 (CH), 43.36 (C), 49.40 (CH), 55.01 (CH), 56.07 (CH), 68.34 (CH₂), 71.04 (CH), 117.44 (CH), 139.56 (C) ppm. MS (70 eV): m/z (%) = 402 (100) [M⁺], 387 (30), 273 (15), 255 (46), 231 (16), 229 (11), 213 (15), 161 (11). HRMS: m/z calcd. for C₂₇H₄₆O₂ [M⁺]: 402.3498, found: 402.3490.

Crystallographic Data for Compound 16: $C_{27}H_{46}O_2$, M = 402.64 gmol⁻¹, crystal size: $0.27 \times 0.12 \times 0.10$ mm³, orthorhombic, space



group $P2_12_12_1$, a = 34.899(7), b = 9.3865(19), c = 7.5000(15) Å, V = 2456.8(9) Å³, Z = 4, $\rho_{calcd.} = 1.089$ g cm⁻³, $\mu = 0.066$ mm⁻¹, $\lambda = 0.71073$ Å, T = 223(2) K, θ range = $3.19-25.40^{\circ}$, reflections collected: 35264, independent: 2611 ($R_{int} = 0.0748$), data/restraints/ parameters: 1947/0/266. The structure was solved by direct methods and refined by full-matrix least-squares on F^2 ; final R indices $[I > 2\sigma(I)]$: $R_1 = 0.0468$; $wR_2 = 0.1065$; maximal residual electron density: 0.242 e Å⁻³. CCDC-697683 contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

(25S)-Δ⁷-Dafachronic Acid [(25S)-3-Keto-5α-cholest-7-en-26-oic Acid] (1): A freshly prepared solution of Jones reagent (CrO₃: 156 mg, 1.565 mmol; concd. H₂SO₄: 137 µL, 2.46 mmol) in water (1 mL) was added to a solution of the diol 16 (126 mg, 313 µmol) in acetone (12 mL) at 0 °C. The resulting mixture was stirred at 0 °C for 90 min, then 2-propanol (5 mL) and water (50 mL) were added. The resulting dark green mixture was extracted with ethyl acetate $(3 \times 50 \text{ mL})$. The combined organic layers were dried with magnesium sulfate and the solvent was evaporated. The residue was purified by flash chromatography on silica gel (petroleum ether/ ethyl acetate, 3:1 + 1% acetic acid) to provide (25*S*)- Δ^7 -dafachronic acid (1), yield 116 mg (89%). Colourless solid; m.p. 139-143 °C (ref.^[5] 143 °C). $[a]_D^{20} = +33.9$ (c = 0.49, CHCl₃). IR (ATR): $\tilde{v} =$ 2939, 2872, 2804, 1724, 1705, 1439, 1420, 1383, 1254, 1233, 1208, 1184, 1159, 1143, 1122, 1013, 974, 938, 844, 832, 793, 749, 729 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): $\delta = 0.54$ (s, 3 H), 0.91 (d, J =6.5 Hz, 3 H), 1.00 (s, 3 H), 1.02–1.06 (m, 1 H), 1.17 (d, J = 7.0 Hz, 3 H), 1.19–1.89 (m, 20 H), 2.03 (dt, J = 12.6, 3.4 Hz, 1 H), 2.12 (ddd, J = 13.4, 6.0, 2.5 Hz, 1 H), 2.20–2.28 (m, 3 H), 2.40 (dd, J = 14.6, 6.0 Hz, 1 H), 2.45 (m, 1 H), 5.17 (d, J = 2.3 Hz, 1 H), 11.12 (br. s, 1 H) ppm. ¹³C NMR and DEPT (125 MHz, CDCl₃): δ = 11.89 (CH₃), 12.45 (CH₃), 17.01 (CH₃), 18.74 (CH₃), 21.68 (CH₂), 22.92 (CH₂), 23.79 (CH₂), 27.92 (CH₂), 30.04 (CH₂), 34.01 (CH₂), 34.38 (C), 35.68 (CH₂), 36.04 (CH), 38.11 (CH₂), 38.75 (CH₂), 39.36 (CH), 39.40 (CH₂), 42.83 (CH), 43.35 (C), 44.22 (CH₂), 48.81 (CH), 54.90 (CH), 56.02 (CH), 117.00 (CH), 139.48 (C), 182.55 (C=O), 212.17 (C=O) ppm. ESI-MS: $m/z = 415.3 [(M + H)^+]$. C₂₇H₄₂O₃ (414.62): C 78.21, H 10.21, found: C 78.21, H 10.31%.

(25S)-26-(Pivaloyloxy)cholest-5-en-3β-ol (17): A 1.0 M solution of tetrabutylammonium fluoride in THF (1.25 mL, 1.25 mmol) was added to a solution of the silvl ether 11 (500 mg, 0.832 mmol) in THF (20 mL). The mixture was heated under reflux for 17 h. After cooling to room temperature, water (50 mL) was added and the mixture was extracted with diethyl ether $(3 \times 50 \text{ mL})$. The combined organic layers were dried with magnesium sulfate and the solvent was evaporated. Purification of the residue by flash chromatography on silica gel (petroleum ether/ethyl acetate, 6:1) provided the alcohol 17, yield 373 mg (92%). Colourless solid; m.p. 101–103 °C. IR (ATR): $\tilde{v} = 3416, 2962, 2934, 2903, 2865, 1729,$ 1479, 1461, 1397, 1376, 1365, 1283, 1160, 1057, 1023, 984, 957, 926, 841, 799, 770, 742 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): δ = 0.66 (s, 3 H), 0.90 (d, J = 7.6 Hz, 3 H), 0.91 (d, J = 6.9 Hz, 3 H), 0.93-1.30 (m, 10 H), 0.99 (s, 3 H), 1.19 (s, 9 H), 1.31-1.59 (m, 10 H), 1.75-1.85 (m, 4 H), 1.94-2.00 (m, 2 H), 2.20-2.30 (m, 2 H), 3.51 (m, 1 H), 3.82 (dd, J = 10.7, 6.7 Hz, 1 H), 3.93 (dd, J = 10.7, 5.6 Hz, 1 H), 5.34 (m, 1 H) ppm. ¹³C NMR and DEPT (125 MHz, CDCl₃): δ = 11.83 (CH₃) 17.07 (CH₃), 18.69 (CH₃), 19.38 (CH₃), 21.05 (CH₂), 23.21 (CH₂), 24.26 (CH₂), 27.21 (3 CH₃), 28.22 (CH₂), 31.63 (CH₂), 31.87 (CH, CH₂), 32.69 (CH), 33.85 (CH₂), 35.67 (CH), 36.14 (CH₂), 36.48 (C), 37.22 (CH₂), 38.84 (C), 39.73 (CH₂), 42.27 (CH₂), 42.29 (C), 50.08 (CH), 56.01 (CH), 56.72 (CH), 69.11 (CH₂), 71.77 (CH), 121.67 (CH), 140.75 (C), 178.66 (C=O) ppm. ESI-MS: $m/z = 509.4 [(M + Na)^+]$. $C_{32}H_{54}O_3$ (486.77): C 78.96, H 11.18, found: C 79.02, H 10.95%.

(25S)-26-(Pivalovloxy)cholest-4-en-3-one (18): Aluminium isopropoxide (152 mg, 746 µmol) was added to a solution of the alcohol 17 (242 mg, 497 µmol) in acetone (1 mL) and toluene (9 mL). The resulting mixture was stirred at 100 °C for 5 h. After cooling to room temperature, water (50 mL) and diethyl ether (50 mL) were added, and the layers were separated. The aqueous layer was extracted with diethyl ether $(2 \times 50 \text{ mL})$ and the combined organic layers were dried with magnesium sulfate. Evaporation of the solvent and purification of the residue by flash chromatography on silica gel (petroleum ether/diethyl ether, 4:1) provided the enone 18, yield 207 mg (86%). Pale yellow solid; m.p. 63-64 °C. IR (ATR): v = 2933, 2868, 2851, 1727, 1671, 1616, 1478, 1466, 1433, 1397, 1377, 1332, 1282, 1227, 1157, 1031, 977, 959, 933, 860, 770, 684 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): δ = 0.69 (s, 3 H), 0.85–0.92 (m, 1 H), 0.89 (d, J = 6.6 Hz, 3 H), 0.91 (d, J = 6.7 Hz, 3 H), 0.93–1.62 (m, 17 H), 1.16 (s, 3 H), 1.19 (s, 9 H), 1.68 (dt, J = 4.8, 14.0 Hz, 1 H), 1.74-1.84 (m, 3 H), 1.98-2.03 (m, 2 H), 2.25 (ddd, J = 14.5, 4.0, 2.3 Hz, 1 H), 2.32 (dt, J = 16.7, 3.8 Hz, 1 H), 2.36–2.44 (m, 2 H), 3.82 (dd, J = 10.7, 6.8 Hz, 1 H), 3.93 (dd, J = 10.7, 5.6 Hz, 1 H),5.71 (s, 1 H) ppm. ¹³C NMR and DEPT (125 MHz, CDCl₃): δ = 11.92 (CH₃), 17.06 (CH₃), 17.35 (CH₃), 18.61 (CH₃), 20.98 (CH₂), 23.21 (CH₂), 24.14 (CH₂), 27.21 (3 CH₃), 28.15 (CH₂), 32.00 (CH₂), 32.69 (CH), 32.92 (CH₂), 33.83 (CH₂), 33.96 (CH₂), 35.57 (CH), 35.64 (CH, CH₂), 36.07 (CH₂), 38.57 (C), 38.83 (C), 39.57 (CH₂), 42.36 (C), 53.76 (CH), 55.82 (CH), 55.96 (CH), 69.08 (CH₂), 123.72 (CH), 171.72 (C), 178.64 (C=O), 199.69 (C=O) ppm. GC-MS $(70 \text{ eV}): m/z \ (\%) = 484 \ (88) \ [M^+], 469 \ (10), 442 \ (17), 382 \ (10), 361$ $(23), 271 (25), 245 (12), 244 (13), 229 (45), 124 (100). C_{32}H_{52}O_{3}$ (484.75): C 79.29, H 10.81, found: C 79.23, H 10.79%.

(25S)-26-Hydroxycholest-4-en-3-one (19): Sodium methoxide (43 mg, 792 $\mu mol)$ was added to a solution of compound 18(192 mg, 396 µmol) in methanol (3 mL). The mixture was stirred at room temperature for 5 d, then 2 N HCl (20 mL) and ethyl acetate (50 mL) were added. The layers were separated and the aqueous layer was extracted with ethyl acetate (2×50 mL). The combined organic layers were dried with magnesium sulfate and the solvent was evaporated. Purification of the residue by flash chromatography on silica gel (petroleum ether/ethyl acetate, 5:1 to 2:1) afforded recovered pivalate 18, yield 29 mg (15%) and the more polar alcohol 19, yield 89 mg (56%). Colourless solid; m.p. 126-127 °C. IR (ATR): $\tilde{v} = 3414$, 2932, 2866, 2850, 1659, 1612, 1462, 1446, 1376, 1332, 1273, 1231, 1190, 1043, 956, 933, 864, 686 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): $\delta = 0.69$ (s, 3 H), 0.86–0.96 (m, 1 H), 0.896 (d, J = 6.5 Hz, 3 H), 0.903 (d, J = 6.7 Hz, 3 H), 0.97-1.14(m, 8 H), 1.16 (s, 3 H), 1.22-1.29 (m, 1 H), 1.32-1.63 (m, 9 H), 1.67 (dt, J = 4.8, 14.0 Hz, 1 H), 1.78–1.86 (m, 2 H), 1.98–2.06 (m, 2 H), 2.25 (ddd, J = 14.6, 4.1, 2.4 Hz, 1 H), 2.32 (dt, J = 17.0, 3.8 Hz, 1 H), 2.34-2.44 (m, 2 H), 3.40 (dd, J = 10.5, 6.5 Hz, 1 H), 3.49 (dd, J = 10.5, 5.7 Hz, 1 H), 5.71 (s, 1 H) ppm. ¹³C NMR and DEPT (125 MHz, CDCl₃): $\delta = 11.92$ (CH₃), 16.70 (CH₃), 17.34 (CH₃), 18.63 (CH₃), 20.98 (CH₂), 23.41 (CH₂), 24.13 (CH₂), 28.16 (CH₂), 32.00 (CH₂), 32.92 (CH₂), 33.61 (CH₂), 33.95 (CH₂), 35.56 (CH), 35.64 (CH₂), 35.71 (CH), 35.79 (CH), 36.15 (CH₂), 38.57 (C), 39.58 (CH₂), 42.35 (C), 53.76 (CH), 55.82 (CH), 56.01 (CH), 68.29 (CH₂), 123.70 (CH), 171.78 (C), 199.74 (C=O) ppm. MS $(70 \text{ eV}): m/z \ (\%) = 400 \ (100) \ [M^+], 385 \ (12), 366 \ (12), 358 \ (31), 277$ (25), 276 (11), 271 (13), 229 (59), 124 (91). HRMS: m/z calcd. for C₂₇H₄₄O₂ [M⁺]: 400.3341, found: 400.3329.

(25*S*)- Δ^4 -Dafachronic Acid [(25*S*)-3-Ketocholest-4-en-26-oic Acid] (2): A freshly prepared solution of Jones reagent (CrO₃: 111 mg,

1.11 mmol; concd. H₂SO₄: 97 µL, 1.743 mmol) in water (0.5 mL) was added to a solution of the alcohol 19 (89 mg, 222 µmol) in acetone (12 mL) at 0 °C. The resulting mixture was stirred at 0 °C for 90 min, then 2-propanol (5 mL) and water (50 mL) were added. The resulting dark green mixture was extracted with ethyl acetate $(3 \times 50 \text{ mL})$. The combined organic layers were dried with magnesium sulfate and the solvent was evaporated. The residue was purified by flash chromatography on silica gel (petroleum ether/ethyl acetate, 2:1) to provide (25S)- Δ^4 -dafachronic acid (2), yield 60 mg (65%). Colourless solid; m.p. 173–174 °C (ref.^[6] 172–175 °C). [a]_D²⁰ = +61.9 (c = 0.47, CHCl₃). IR (ATR): \tilde{v} = 2929, 2848, 1725, 1649, 1610, 1467, 1450, 1413, 1375, 1362, 1333, 1306, 1282, 1237, 1199, 1168, 1117, 1030, 947, 932, 901, 871, 843, 781 cm⁻¹. ¹H NMR $(500 \text{ MHz}, \text{CDCl}_3)$: $\delta = 0.69 \text{ (s, 3 H)}, 0.82-0.93 \text{ (m, 1 H)}, 0.89 \text{ (d,})$ J = 6.5 Hz, 3 H), 0.96–1.29 (m, 8 H), 1.16 (s, 3 H), 1.17 (d, J =6.9 Hz, 3 H), 1.31–1.69 (m, 9 H), 1.68 (dt, J = 4.7, 13.9 Hz, 1 H), 1.78-1.86 (m, 2 H), 1.98-2.03 (m, 2 H), 2.25 (ddd, J = 14.6, 4.0,2.4 Hz, 1 H), 2.32 (dt, J = 17.3, 4.0 Hz, 1 H), 2.36–2.44 (m, 2 H), 2.45 (sext, J = 6.9 Hz, 1 H), 5.71 (s, 1 H) ppm. ¹³C NMR and DEPT (125 MHz, CDCl₃): δ = 11.93 (CH₃), 17.01 (CH₃), 17.35 (CH₃), 18.54 (CH₃), 20.99 (CH₂), 23.69 (CH₂), 24.14 (CH₂), 28.15 (CH₂), 32.00 (CH₂), 32.93 (CH₂), 33.94 (CH₂), 34.00 (CH₂), 35.58 (2 CH), 35.64 (CH₂), 35.68 (CH₂), 38.58 (C), 39.34 (CH), 39.58 (CH₂), 42.37 (C), 53.75 (CH), 55.82 (CH), 55.98 (CH), 123.71 (CH), 171.86 (C), 182.35 (C=O), 199.84 (C=O) ppm. ESI-MS m/z = 415.3 $[(M + H)^+]$. C₂₇H₄₂O₃ (414.62): C 78.21, H 10.21, found: C 78.21, H 10.12%.

(25S)-26-Hydroxycholesterol [(25S)-Cholest-5-en-3ß,26-diol] (20): A 1.0 M solution of tetrabutylammonium fluoride in THF (249 µL, 249 µmol) was added to a solution of compound 11 (100 mg, 166 µmol) in THF (10 mL). The mixture was heated under reflux for 17 h. After cooling to room temperature, water (50 mL) was added and the resulting mixture was extracted with diethyl ether $(3 \times 50 \text{ mL})$. The combined organic layers were dried with magnesium sulfate and the solvent was evaporated. The crude product was dissolved in THF (15 mL) and lithium aluminium hydride (25 mg, 664 µmol) was added. The mixture was stirred at room temperature for 16 h, then water (10 mL) and 10% HCl were added (10 mL). The mixture was extracted with ethyl acetate $(3 \times 50 \text{ mL})$ and the combined organic layers were dried with magnesium sulfate. Evaporation of the solvent and purification of the residue by flash chromatography on silica gel (petroleum ether/ethyl acetate, 3:1) provided the diol 20, yield 62 mg (93%). Colourless solid; m.p. 173–175 °C (ref.^[22] 171–174 °C). IR (ATR): v = 3318, 2931, 2864, 1464, 1376, 1231, 1193, 1132, 1107, 1053, 1039, 1022, 987, 954, 926, 839, 799, 736 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): δ = 0.66 (s, 3 H), 0.88-1.17 (m, 9 H), 0.90 (d, J = 6.8 Hz, 6 H), 0.99 (s, 3 H), 1.20–1.27 (m, 1 H), 1.33–1.63 (m, 11 H), 1.77–1.85 (m, 3 H), 1.93– 2.01 (m, 2 H), 2.23 (m, 1 H), 2.28 (ddd, J = 13.0, 5.0, 1.9 Hz, 1 H), 3.40 (dd, J = 10.5, 6.5 Hz, 1 H), 3.48–3.54 (m, 1 H), 3.50 (dd, J = 10.5, 5.7 Hz, 1 H), 5.33 (m, 1 H) ppm. ¹³C NMR and DEPT $(125 \text{ MHz}, \text{ CDCl}_3): \delta = 11.84 \text{ (CH}_3), 16.70 \text{ (CH}_3), 18.71 \text{ (CH}_3),$ 19.38 (CH₃), 21.04 (CH₂), 23.42 (CH₂), 24.26 (CH₂), 28.22 (CH₂), 31.61 (CH₂), 31.86 (CH), 31.87 (CH₂), 33.63 (CH₂), 35.75 (CH), 35.80 (CH), 36.22 (CH₂), 36.47 (C), 37.21 (CH₂), 39.73 (CH₂), 42.26 (CH₂), 42.28 (C), 50.07 (CH), 56.06 (CH), 56.72 (CH), 68.32 (CH₂), 71.76 (CH), 121.68 (CH), 140.73 (C) ppm. MS (70 eV): m/z $(\%) = 402 (100) [M^+], 387 (38), 384 (70), 369 (43), 317 (47), 291$ (76), 273 (24), 255 (32), 231 (20), 213 (42). HRMS: m/z calcd. for C₂₇H₄₆O₂ [M⁺]: 402.3498, found: 402.3494. C₂₇H₄₆O₂ (402.65): C 80.54, H 11.51, found: C 80.71, H 11.59%.

(25S)-5a-Cholestan-3 β ,26-diol (21): A solution of the diol 20 (127 mg, 315 μ mol) in dichloromethane (5 mL) was added to a mix-

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The reaction mixture was stirred under an hydrogen atmosphere at room temperature for 24 h and then filtered with ethyl acetate over a short pad of Celite[®]. The solvent was evaporated to provide the pure saturated diol 21, yield 126 mg (99%). Colourless solid; m.p. 169–171 °C. IR (ATR): $\tilde{v} = 3240, 2969, 2932, 2848, 1463, 1449,$ 1385, 1369, 1358, 1332, 1320, 1256, 1236, 1171, 1127, 1080, 1049, 1030, 1005, 987, 956, 917, 797, 736, 719 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): $\delta = 0.58-0.61$ (m, 1 H), 0.63 (s, 3 H), 0.79 (s, 3 H), 0.80-0.93 (m, 1 H), 0.89 (d, J = 6.6 Hz, 3 H), 0.90 (d, J = 6.7 Hz, 3 H),0.94-1.15 (m, 9 H), 1.17-1.49 (m, 12 H), 1.51-1.66 (m, 4 H), 1.69 (dt, J = 13.2, 3.6 Hz, 1 H), 1.75-1.81 (m, 2 H), 1.94 (dt, J = 12.6,3.4 Hz, 1 H), 3.40 (dd, J = 10.5, 6.5 Hz, 1 H), 3.50 (dd, J = 10.5, 5.7 Hz, 1 H), 3.57 (m, 1 H) ppm. ¹³C NMR and DEPT (125 MHz, $CDCl_3$): $\delta = 12.06 (CH_3), 12.31 (CH_3), 16.71 (CH_3), 18.67 (CH_3),$ 21.23 (CH₂), 23.43 (CH₂), 24.19 (CH₂), 28.25 (CH₂), 28.71 (CH₂), 31.50 (CH₂), 32.07 (CH₂), 33.64 (CH₂), 35.44 (C), 35.48 (CH), 35.77 (CH), 35.81 (CH), 36.22 (CH₂), 36.98 (CH₂), 38.19 (CH₂), 40.01 (CH₂), 42.57 (C), 44.83 (CH), 54.32 (CH), 56.21 (CH), 56.46 (CH), 68.35 (CH₂), 71.37 (CH) ppm. MS (70 eV): m/z (%) = 404 (100) [M⁺], 389 (22), 388 (14), 386 (16), 371 (25), 278 (11), 248 (22), 234 (40), 233 (90), 217 (52), 215 (78). HRMS: m/z calcd. for $C_{27}H_{48}O_2\ [M^+]$: 404.3654, found: 404.3652. $C_{27}H_{48}O_2\ (404.67)$: C 80.14, H 11.96, found: C 79.53, H 11.66%.

(25S)-Dafachronic Acid [(25S)-3-Keto-5α-cholestan-26-oic Acid] (3): A freshly prepared solution of Jones reagent (CrO₃: 124 mg, 1.24 mmol; concd. H₂SO₄: 111 µL, 1.99 mmol) in water (0.7 mL) was added to a solution of the diol 21 (100 mg, 247 µmol) in acetone (12 mL) at 0 °C. The resulting mixture was stirred at 0 °C for 60 min, then 2-propanol (5 mL) and water (50 mL) were added. The resulting dark green mixture was extracted with ethyl acetate $(3 \times 50 \text{ mL})$. The combined organic layers were dried with magnesium sulfate and the solvent was evaporated. The residue was purified by flash chromatography on silica gel (petroleum ether/ethyl acetate, 3:1 + 1% acetic acid) to provide (25S)-dafachronic acid (3), yield 91 mg (88%). Light yellow solid; m.p. 123-126 °C. $[a]_{D}^{20} = +44.9 \ (c = 1.0, \text{CHCl}_3)$. IR (ATR): $\tilde{v} = 2931, 2863,$ 1702, 1443, 1416, 1379, 1292, 1232, 1173, 1030, 955, 804, 732 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): δ = 0.66 (s, 3 H), 0.68–0.73 (m, 1 H), 0.84–0.92 (m, 1 H), 0.88 (d, *J* = 6.5 Hz, 3 H), 0.97–1.41 (m, 16 H), 0.99 (s, 3 H), 1.17 (d, J = 7.0 Hz, 3 H), 1.47–1.58 (m, 3 H), 1.63-1.70 (m, 2 H), 1.76-1.83 (m, 1 H), 1.95-2.02 (m, 2 H), 2.07 (ddd, J = 14.7, 3.8, 2.2 Hz, 1 H), 2.24 (d, J = 14.7 Hz, 1 H), 2.26–2.30 (m, 1 H), 2.35 (dd, J = 13.9, 6.5 Hz, 1 H), 2.45 (sext, J = 7.0 Hz, 1 H) ppm. ¹³C NMR and DEPT (125 MHz, CDCl₃): δ = 11.45 (CH₃), 12.05 (CH₃), 16.99 (CH₃), 18.56 (CH₃), 21.42 (CH₂), 23.71 (CH₂), 24.19 (CH₂), 28.21 (CH₂), 28.94 (CH₂), 31.68 (CH₂), 34.00 (CH₂), 35.36 (CH), 35.61 (C, CH), 35.71 (CH₂), 38.18 (CH₂), 38.53 (CH₂), 39.37 (CH), 39.86 (CH₂), 42.57 (C), 44.71 (CH₂), 46.67 (CH), 53.74 (CH), 56.15 (CH), 56.23 (CH), 182.70 (C=O), 212.41 (C=O) ppm. MS (70 eV): m/z (%) = 416 (33) [M⁺], 402 (19), 398 (14), 387 (13), 370 (18), 246 (23), 233 (13), 232 (42), 231 (100), 218 (19), 217 (53). HRMS: m/z calcd. for C₂₇H₄₄O₃ [M⁺]: 416.3290, found: 416.3291.

(25*S*)-3β-(*tert*-Butyldimethylsilyloxy)cholest-5-en-26-ol (22): Lithium aluminium hydride (63 mg, 1.664 mmol) was slowly added to a solution of the pivalate 11 (250 mg, 0.416 mmol) in THF (10 mL) at 0 °C. The reaction mixture was stirred at room temperature for 17 h, then water (10 mL) and 10% sulfuric acid were added. The mixture was extracted with diethyl ether (3×100 mL) and the combined organic layers were dried with magnesium sulfate. Removal of the solvent and purification of the residue by flash chromatog-



raphy on silica gel (petroleum ether/diethyl ether, 5:1) provided the alcohol 22, yield 195 mg (91%). Colourless solid; m.p. 165-166 °C. IR (ATR): $\tilde{v} = 3302, 2929, 2857, 1462, 1381, 1367, 1250, 1196,$ 1093, 1024, 1006, 989, 958, 886, 869, 835, 803, 775, 732, 669 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): δ = 0.04 (s, 6 H), 0.66 (s, 3 H), 0.87– 0.94 (m, 1 H), 0.88 (s, 9 H), 0.90 (d, J = 6.6 Hz, 3 H), 0.91 (d, J = 6.7 Hz, 3 H), 0.95-1.17 (m, 8 H), 0.98 (s, 3 H), 1.24 (m, 1 H), 1.33-1.62 (m, 11 H), 1.68–1.72 (m, 1 H), 1.77–1.85 (m, 1 H), 1.79 (dt, J = 13.4, 3.6 Hz, 1 H), 1.92–2.01 (m, 2 H), 2.15 (ddd, J = 13.3, 4.9, 2.2 Hz, 1 H), 2.26 (m, 1 H), 3.41 (dd, J = 10.5, 6.6 Hz, 1 H), 3.47 (m, 1 H), 3.50 (dd, J = 10.5, 5.7 Hz, 1 H), 5.30 (m, 1 H) ppm. ¹³C NMR and DEPT (125 MHz, CDCl₃): $\delta = -4.60$ (2 CH₃), 11.84 (CH₃), 16.72 (CH₃), 18.27 (C), 18.72 (CH₃), 19.42 (CH₃), 21.04 (CH₂), 23.41 (CH₂), 24.27 (CH₂), 25.93 (3 CH₃), 28.24 (CH₂), 31.88 (CH), 31.92 (CH₂), 32.06 (CH₂), 33.65 (CH₂), 35.76 (CH), 35.82 (CH), 36.24 (CH₂), 36.56 (C), 37.36 (CH₂), 39.78 (CH₂), 42.30 (C), 42.79 (CH₂), 50.17 (CH), 56.06 (CH), 56.77 (CH), 68.35 (CH₂), 72.63 (CH), 121.15 (CH), 141.55 (C) ppm. GC-MS (70 eV): m/z $(\%) = 459 (100) [(M - tBu)^+], 389 (17), 377 (54), 343 (10), 331 (19),$ 281 (21), 273 (12), 269 (14). C₃₃H₆₀O₂Si (516.91): C 76.68, H 11.70, found: C 76.82, H 11.88%.

(25S)-3β-(tert-Butyldimethylsilyloxy)cholest-5-en-26-oic Acid (23): Oxalyl chloride (84 µL, 0.994 mmol) was added to a solution of DMSO (141 μ L, 1.988 mmol) in dichloromethane (5 mL) at -78 °C. After 5 min, a solution of the alcohol 22 (257 mg, 0.497 mmol) in dichloromethane (10 mL) was added and the reaction mixture was stirred at -78 °C for 20 min. Then, triethylamine (346 µL, 2.485 mmol) was added dropwise, the solution was warmed to room temperature and stirring was continued for 10 min. A saturated aqueous solution of ammonium chloride (50 mL) was added and the layers were separated. The aqueous layer was extracted with dichloromethane (2×50 mL). The combined organic layers were dried with magnesium sulfate and the solvent was evaporated to afford the aldehyde. 2-Methyl-2-butene $(527 \,\mu\text{L}, 4.97 \,\text{mmol})$ and KH₂PO₄ (200 mg) were added to the solution of the aldehyde in THF (6 mL) and water (1 mL). After addition of a solution of sodium chlorite (90 mg, 0.994 mmol) in water (1 mL), the mixture was stirred at room temperature for 24 h. Then, 10% HCl (25 mL) and diethyl ether (50 mL) were added, and the layers were separated. The aqueous layer was extracted with diethyl ether $(2 \times 50 \text{ mL})$ and the combined organic layers were dried with magnesium sulfate. Removal of the solvent and purification of the residue by flash chromatography on silica gel (petroleum ether/diethyl ether, 3:1) provided the acid 23, yield 235 mg (89%). Colourless solid; m.p. 183–185 °C. IR (ATR): \tilde{v} = 2931, 2880, 2854, 1704, 1464, 1418, 1380, 1294, 1248, 1226, 1198, 1083, 988, 957, 888, 870, 835, 803, 774, 733, 669 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): δ = 0.05 (s, 6 H) 0.65 (s, 3 H), 0.86–0.94 (m, 1 H), 0.88 (s, 9 H), 0.90 (d, J = 6.5 Hz, 3 H), 0.95–1.28 (m, 8 H), 0.98 (s, 3 H), 1.17 (d, J = 7.0 Hz, 3 H), 1.31–1.59 (m, 10 H), 1.63– 1.72 (m, 2 H), 1.76-1.83 (m, 2 H), 1.92-1.98 (m, 1 H), 1.98 (dt, J = 12.6, 3.2 Hz, 1 H), 2.15 (ddd, J = 13.3, 4.9, 2.1 Hz, 1 H), 2.26 (m, 1 H), 2.45 (sext, J = 7.0 Hz, 1 H), 3.47 (m, 1 H), 5.30 (m, 1 H), 11.08 (br. s, 1 H) ppm. ¹³C NMR and DEPT (125 MHz, CDCl₃): δ = -4.60 (2 CH₃), 11.84 (CH₃), 16.99 (CH₃), 18.27 (C), 18.62 (CH₃), 19.41 (CH₃), 21.04 (CH₂), 23.71 (CH₂), 24.27 (CH₂), 25.93 (3 CH₃), 28.22 (CH₂), 31.88 (CH), 31.91 (CH₂), 32.05 (CH₂), 34.03 (CH₂), 35.63 (CH), 35.76 (CH₂), 36.56 (C), 37.36 (CH₂), 39.35 (CH), 39.76 (CH₂), 42.31 (C), 42.79 (CH₂), 50.16 (CH), 56.04 (CH), 56.76 (CH), 72.64 (CH), 121.14 (CH), 141.55 (C), 182.60 (C=O) ppm. ESI-MS: $m/z = 553.4 [(M + Na)^+]$. C₃₃H₅₈O₃Si (530.90): C 74.66, H 11.01, found: C 74.75, H 10.94%.

Methyl (25S)-3β-Hydroxycholesten-5-en-26-oate (24): A catalytic amount of concentrated sulfuric acid was added to a solution of the acid 23 (227 mg, 0.428 mmol) in methanol (10 mL) and the mixture was heated under reflux for 16 h. After cooling to room temperature, a saturated aqueous solution of sodium hydrogen carbonate (25 mL) and ethyl acetate (50 mL) were added. The layers were separated and the aqueous layer was extracted with ethyl acetate (2×50 mL). The combined organic layers were dried with magnesium sulfate and the solvent was evaporated. The residue was purified by flash chromatography on silica gel (petroleum ether/ethyl acetate, 6:1) to provide the methyl ester 24, yield 161 mg (87%). Colourless crystals; m.p. 118-119 °C. IR (ATR): v = 3443, 2931, 2901, 2886, 2866, 1732, 1460, 1378, 1363, 1312, 1222, 1192, 1164, 1140, 1107, 1054, 1041, 1025, 1011, 987, 959, 841, 804, 765, 741 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): $\delta = 0.66$ (s, 3 H), 0.87-1.16 (m, 8 H), 0.89 (d, J = 6.5 Hz, 3 H), 0.99 (s, 3 H), 1.13 (d, J = 7.0 Hz, 3 H), 1.19–1.66 (m, 12 H), 1.76–1.85 (m, 3 H), 1.93–1.99 (m, 1 H), 1.99 (dt, J = 12.6, 3.5 Hz, 1 H), 2.22– 2.24 (m, 1 H), 2.28 (ddd, J = 13.0, 5.1, 2.0 Hz, 1 H), 2.42 (sext, J = 7.0 Hz, 1 H), 3.51 (m, 1 H), 3.66 (s, 3 H), 5.34 (m, 1 H) ppm. ¹³C NMR and DEPT (125 MHz, CDCl₃): $\delta = 11.84$ (CH₃), 17.23 (CH₃), 18.60 (CH₃), 19.38 (CH₃), 21.05 (CH₂), 23.76 (CH₂), 24.26 (CH₂), 28.20 (CH₂), 31.62 (CH₂), 31.86 (CH), 31.87 (CH₂), 34.31 (CH₂), 35.61 (CH), 35.75 (CH₂), 36.47 (C), 37.22 (CH₂), 39.51 (CH), 39.73 (CH₂), 42.27 (CH₂), 42.29 (C), 50.07 (CH), 51.45 (CH₃), 56.02 (CH), 56.72 (CH), 71.77 (CH), 121.68 (CH), 140.74 (C), 177.45 (C=O) ppm. C₂₈H₄₆O₃ (430.66): C 78.09, H 10.77, found: C 78.06, H 10.88%.

(25S)-Cholestenoic Acid [(25S)-3β-Hydroxycholest-5-en-26-oic Acid] (4): Lithium hydroxide (11.5 mg, 480 µmol) was added to a solution of the methyl ester 24 (69 mg, 160 µmol) in THF/methanol/water (1:1:1, 6 mL) and the mixture was stirred at room temperature for 24 h. Methanol and THF were removed in vacuo and the residue was thoroughly extracted with dichloromethane $(3 \times 30 \text{ mL})$ to remove traces of unreacted starting material. The combined dichloromethane layers were discarded. Then, 10% hydrochloric acid was added to the aqueous residue (pH < 4) and the mixture was extracted with ethyl acetate (3×40 mL). The combined organic layers were dried with magnesium sulfate and the solvent was evaporated to provide pure (25S)-cholestenoic acid (4), yield 67 mg (99%). An analytically pure sample was obtained by recrystallization from acetonitrile. Colourless solid; m.p. 172-175 °C (MeCN) (ref.^[6] 157-160 °C). $[a]_{D}^{20} = -22.9 (c = 0.14, CHCl_3)$. IR (ATR): $\tilde{v} = 3262, 2934,$ 2922, 2890, 2862, 1674, 1457, 1434, 1421, 1374, 1277, 1225, 1137, 1113, 1088, 1052, 1020, 987, 953, 927, 890, 841, 819, 797, 737, 658 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): $\delta = 0.66$ (s, 3 H), 0.87–1.27 (m, 9 H), 0.90 (d, J = 6.5 Hz, 3 H), 0.99 (s, 3 H), 1.17 (d, J =7.0 Hz, 3 H), 1.31-1.59 (m, 10 H), 1.63-1.69 (m, 1 H), 1.77-1.85 (m, 3 H), 1.93–2.01 (m, 2 H), 2.22–2.30 (m, 2 H), 2.46 (sext, J =7.0 Hz, 1 H), 3.52 (m, 1 H), 5.34 (m, 1 H) ppm. $^{13}\mathrm{C}$ NMR and DEPT (125 MHz, CDCl₃): δ = 11.84 (CH₃), 17.02 (CH₃), 18.62 (CH₃), 19.38 (CH₃), 21.05 (CH₂), 23.70 (CH₂), 24.26 (CH₂), 28.21 (CH₂), 31.61 (CH₂), 31.87 (CH, CH₂), 34.05 (CH₂), 35.63 (CH), 35.75 (CH₂), 36.48 (C), 37.22 (CH₂), 39.21 (CH), 39.73 (CH₂), 42.24 (CH₂), 42.30 (C), 50.07 (CH), 56.03 (CH), 56.71 (CH), 71.81 (CH), 121.71 (CH), 140.71 (C), 181.58 (C=O) ppm. ESI-MS: m/z = 399.4 $[(M + H - H_2O)^+]$. C₂₇H₄₄O₃ (416.64): C 77.83, H 10.64, found: C 77.99, H 10.77%.

Supporting Information (see also the footnote on the first page of this article): Copies of the ¹H and ¹³C NMR spectra for compound 10 and copies of the HSQC spectra for the (25*S*)-steroidal acids 1-4.

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